

DHA, an essential fatty acid required for neurodevelopment, metabolism in healthy pregnancy and preeclampsia

Dilys J Freeman
Institute for Cardiovascular and Medical Sciences
University of Glasgow



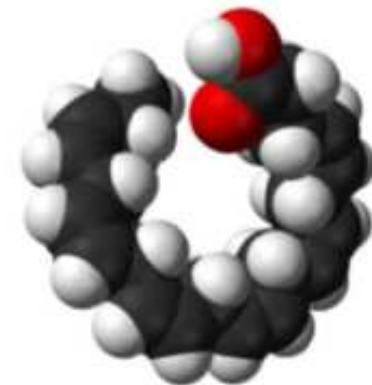
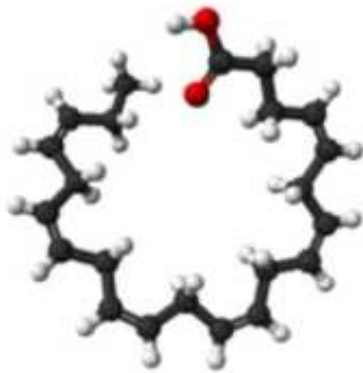
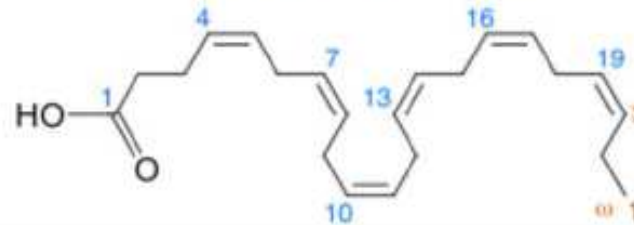


A long chain polyunsaturated fatty acid (LC PUFA) 22:6n-3

Docosahexaenoic acid

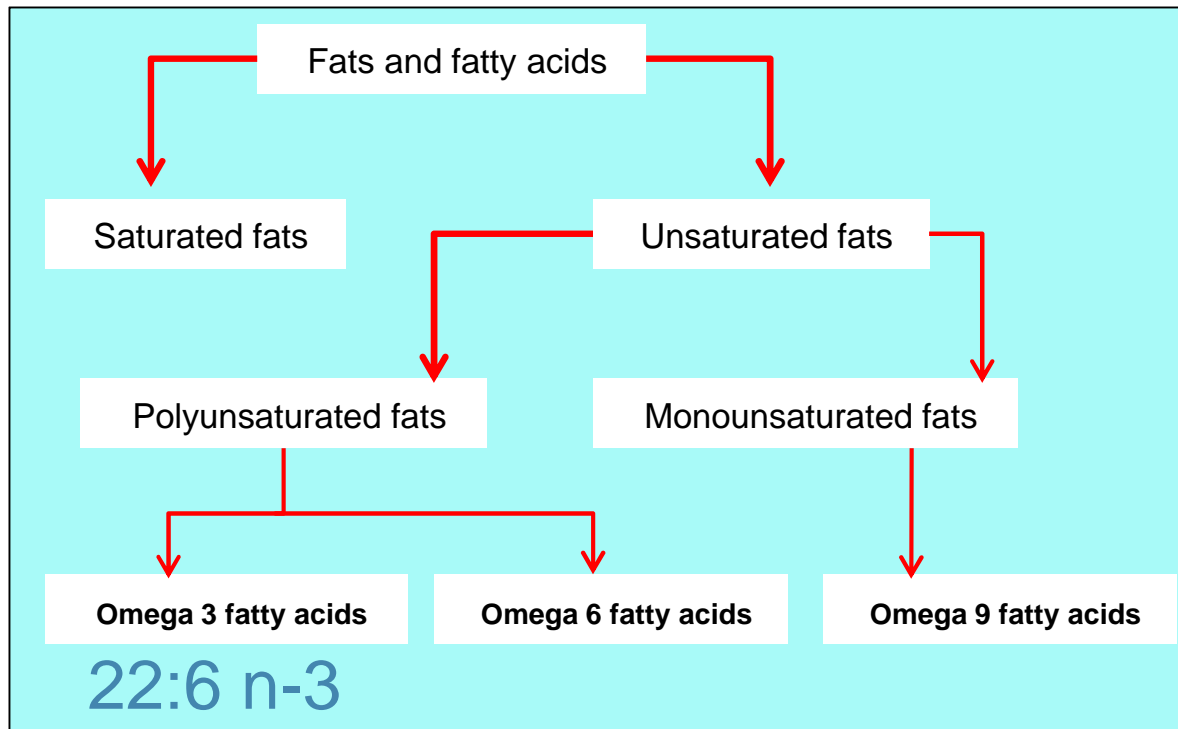
From Wikipedia, the free encyclopedia

Docosahexaenoic acid

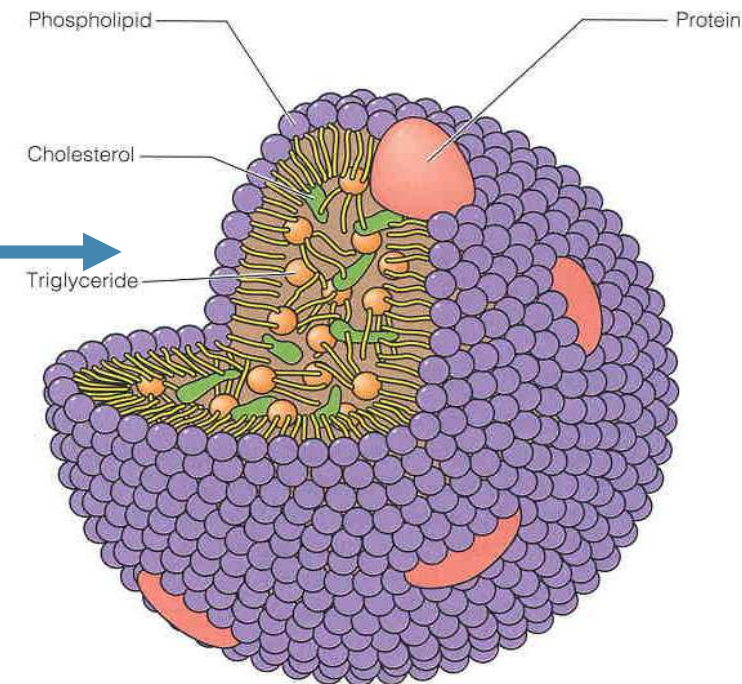


A bit of background

3 x NEFA + glycerol = TG



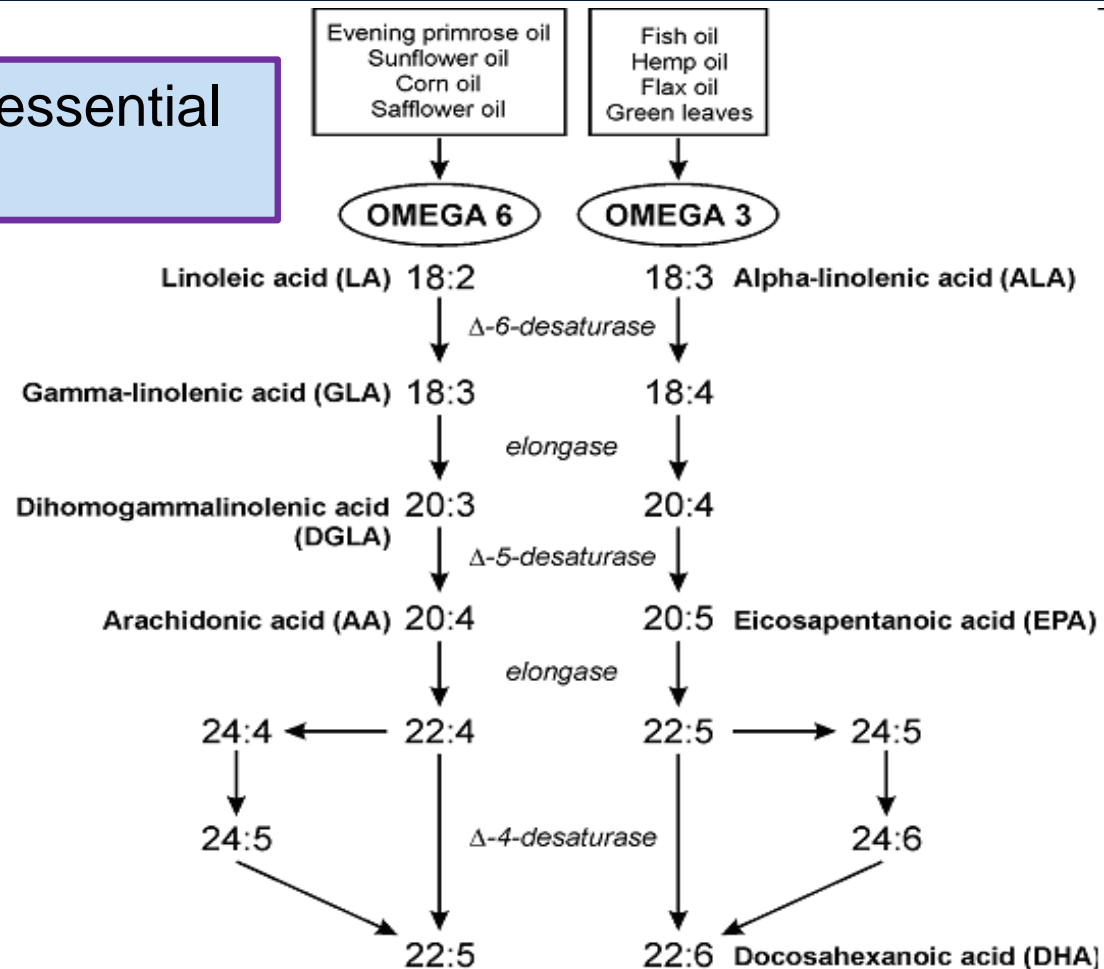
TG carried in lipoprotein core



The main lipoprotein carrier of TG is very low density lipoprotein (VLDL)

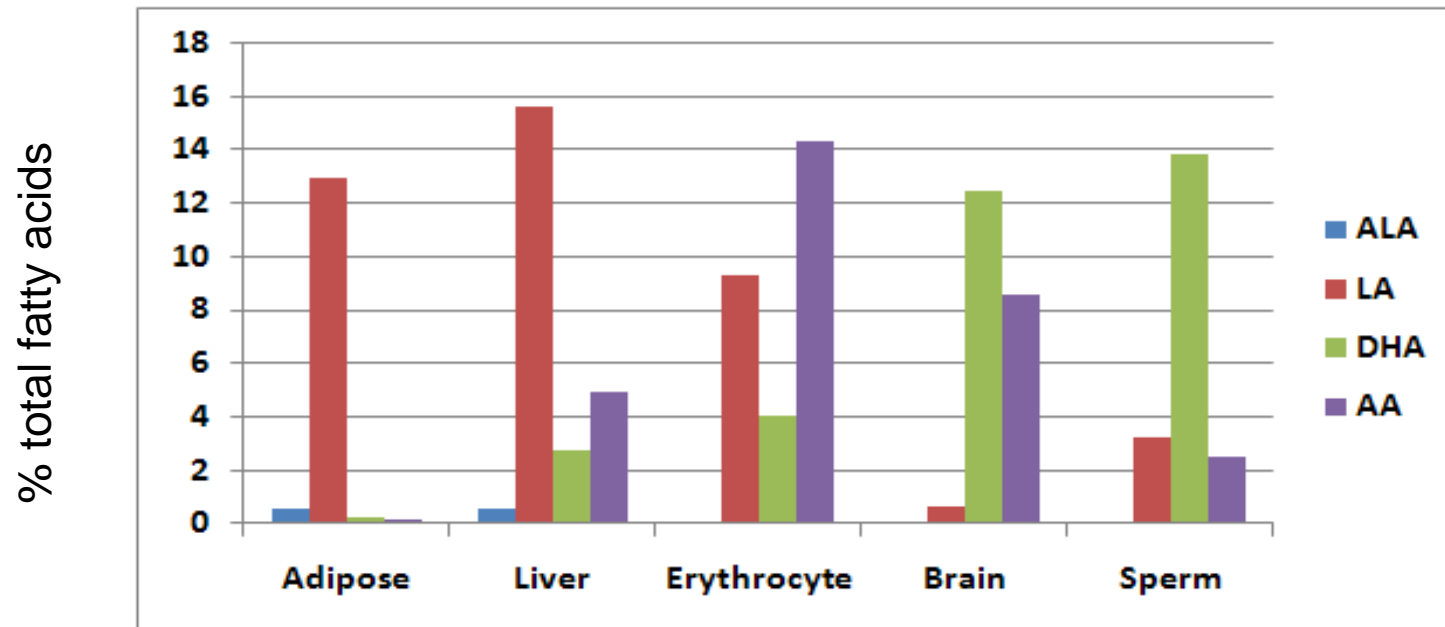
PUFA synthetic pathways

LA and ALA are essential fatty acids



LC-PUFA, especially DHA, are known to be very important for neural development

DHA content of tissues





DHA increases membrane fluidity

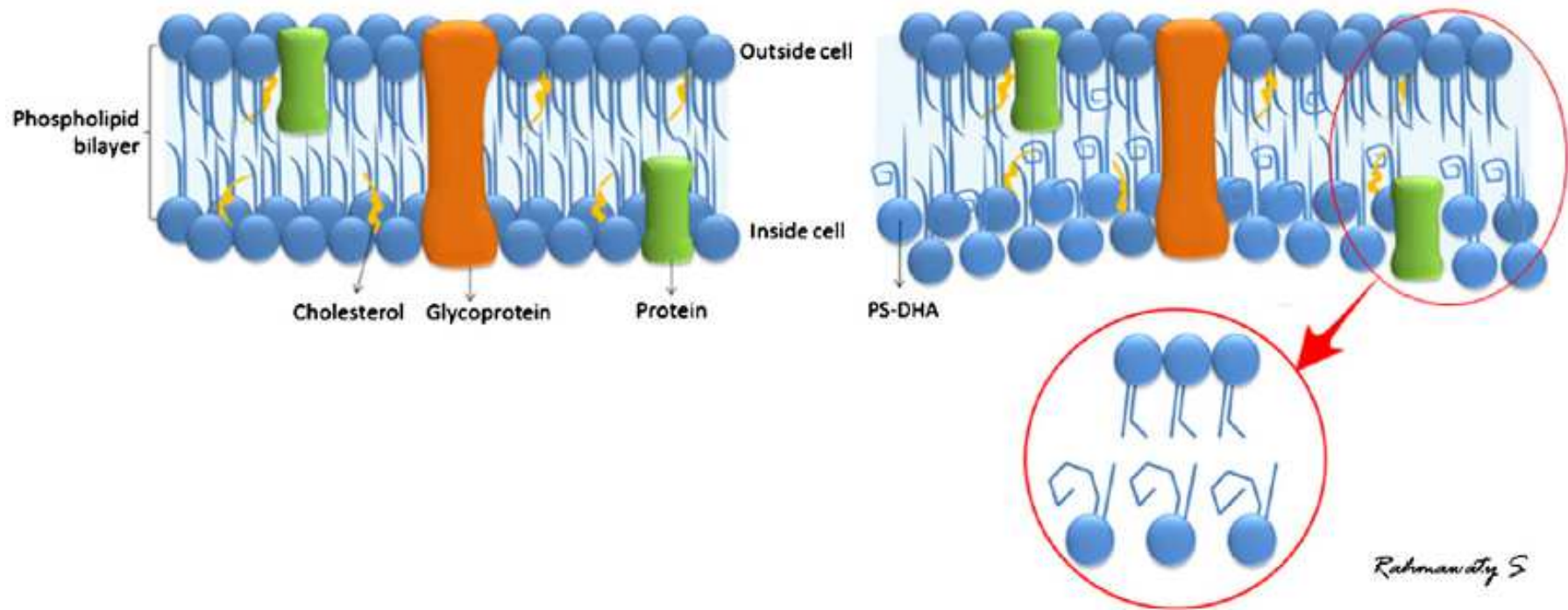


Diagram 3. Phospholipid bilayer DHA.

DHA increases neurone survival

In DHA deficiency, omega-6 DPA (22:5n-6) used in place of DHA in brain phosphatidyl serine and is thought to be a survival mechanism

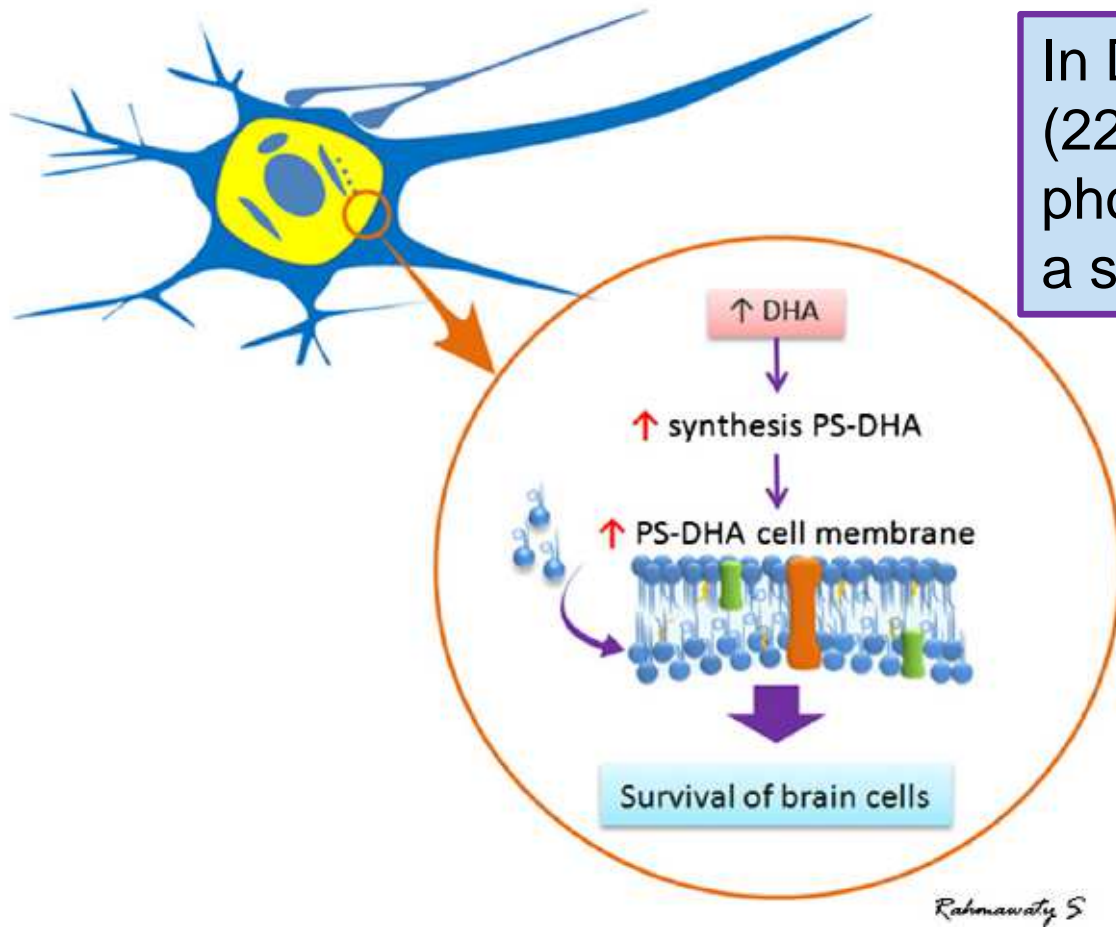


Diagram 6. Neural survival (PS-DHA).

DHA required for neurite outgrowth

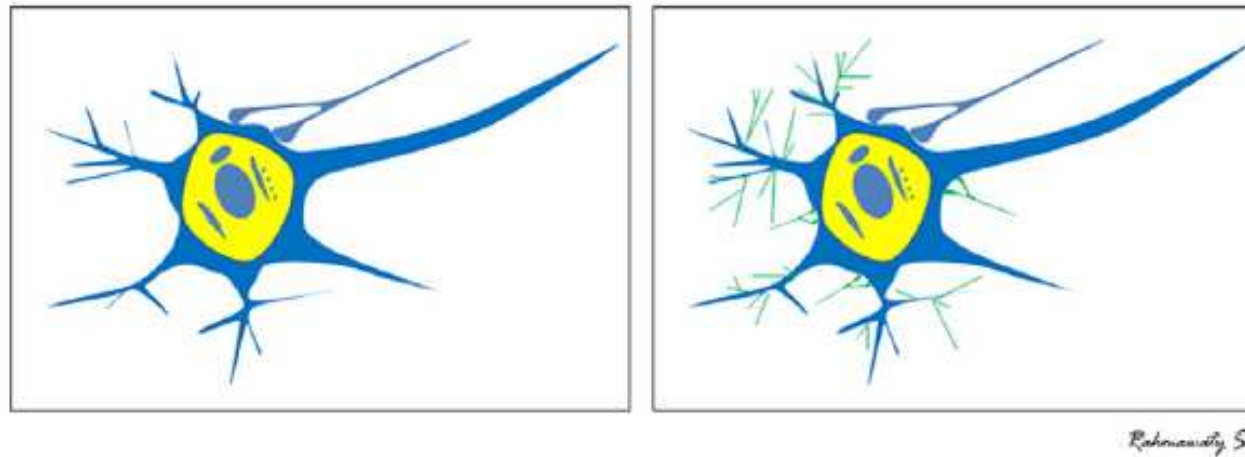
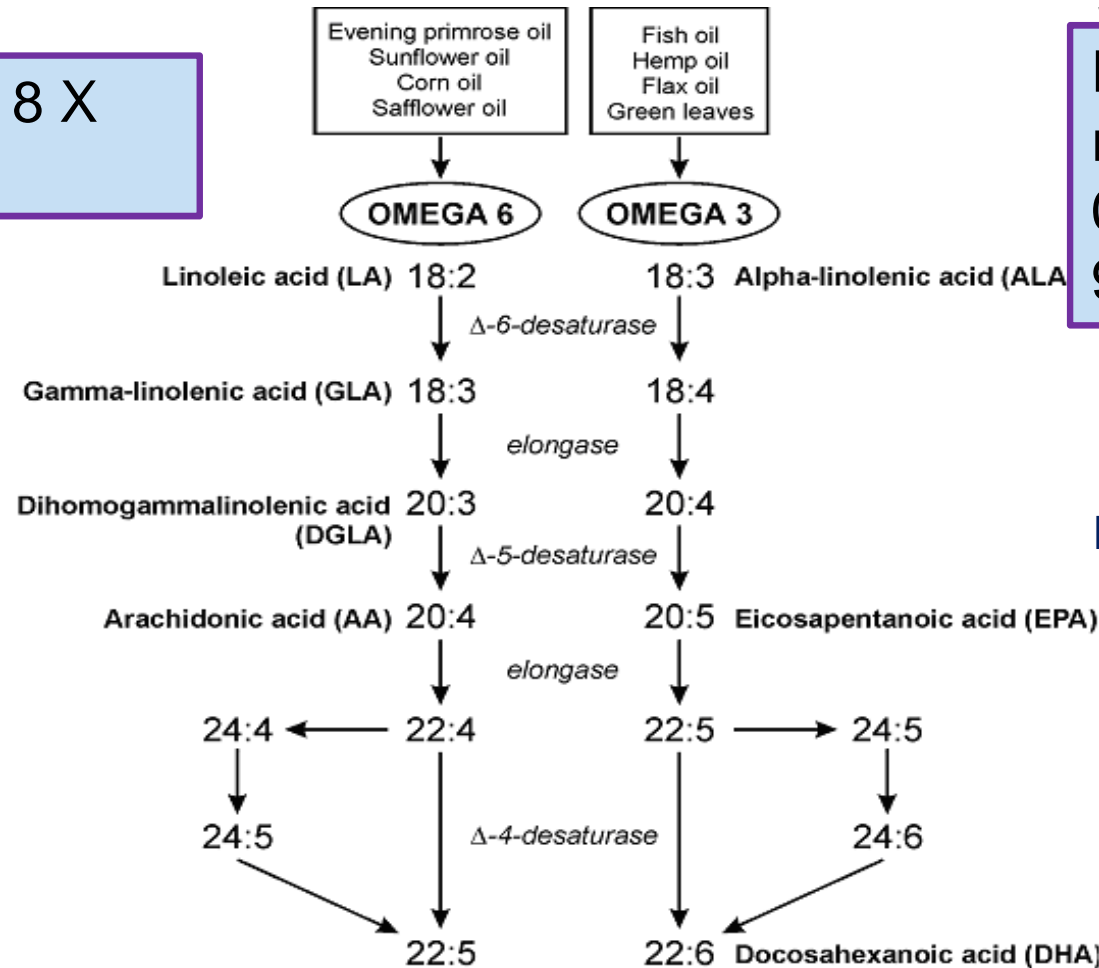


Diagram 2. The effect of DHA on neurite growth.

DHA interaction with the plasma membrane protein syntaxin 3 required for the membrane fusion necessary for neurite outgrowth in developing neurones

Humans are not great at making DHA

Dietary LA intake is 8 X
ALA intake



Fractional conversion
rates of ALA to DHA
0.04% in men
9% in women

Burdge et al Br J Nutr 2002

The critical requirement of DHA for fetal brain development, and the poor efficiency of its synthesis in humans, is therefore a metabolic problem to be overcome in pregnancy!

Fetal plasma is enriched in DHA

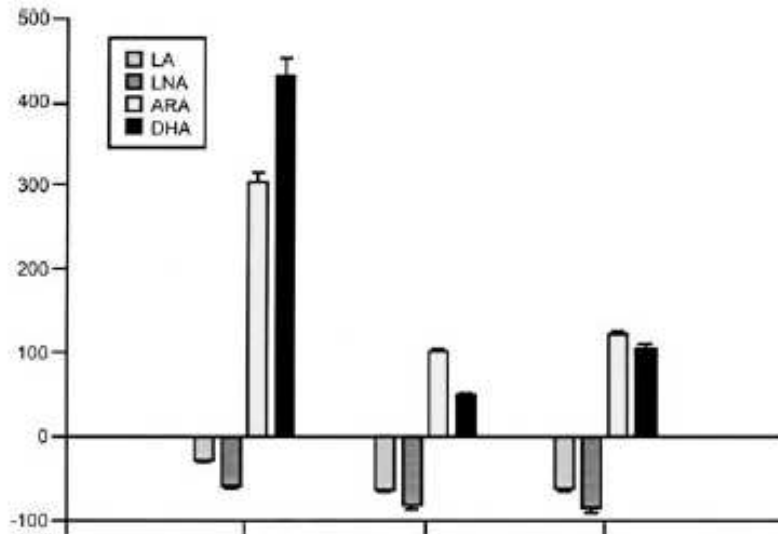
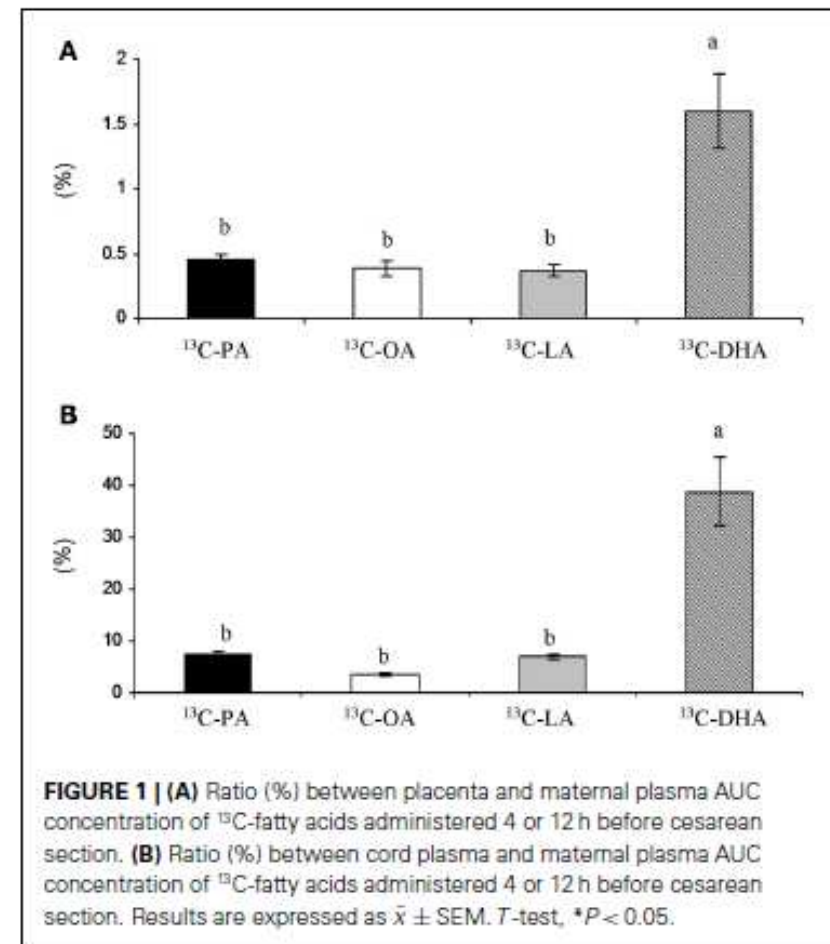


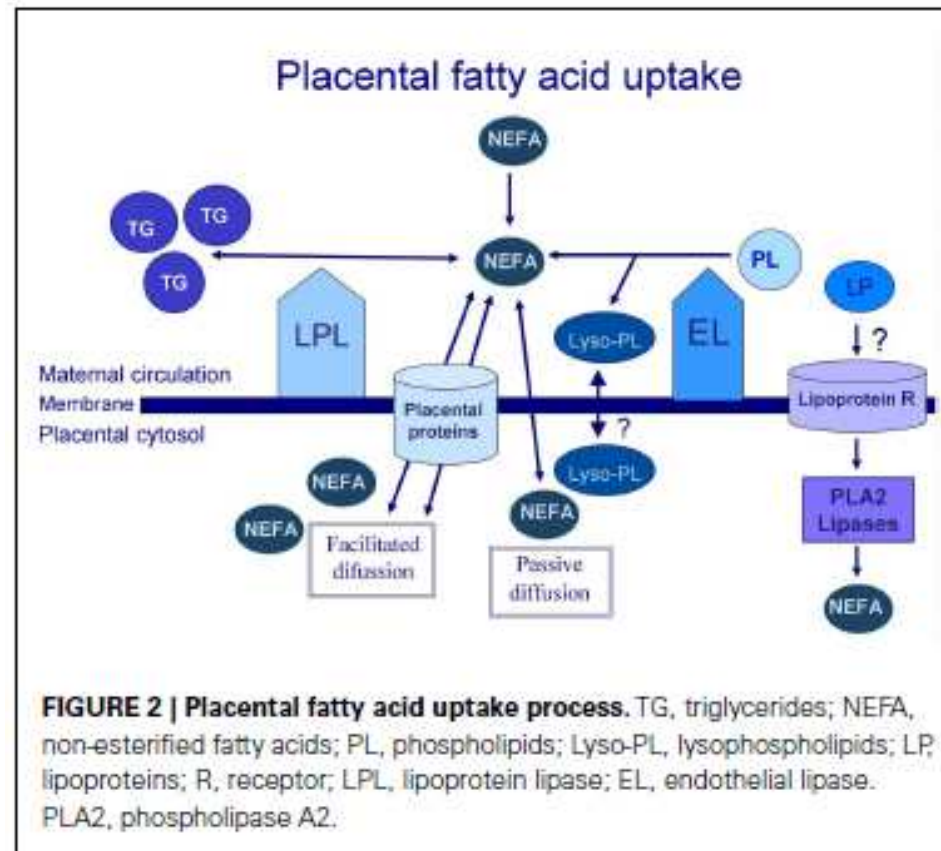
Fig 2. Fatty acid enrichment in fetal compared with maternal plasma. Relative enrichment of LA, LNA, ARA, and DHA in fetal compared with maternal plasma was calculated for each mother-fetal cord plasma pair as the difference in the given fatty acid in the maternal compared to fetal plasma/maternal plasma $\times 100\%$. Values shown are mean \pm SEM, $n = 55$. Adapted from data published in Reference 89.



Placenta preferentially transports DHA

LA , ALA

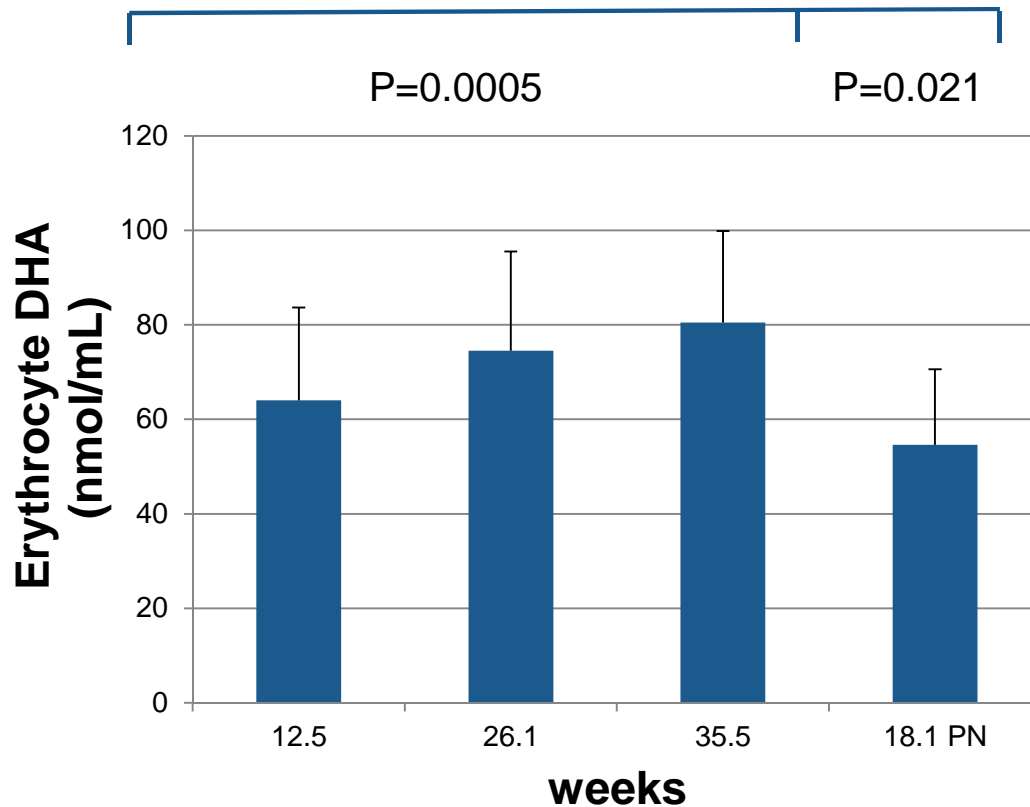
LA , ALA



DHA

DHA

Maternal erythrocyte DHA concentration in pregnancy and post partum



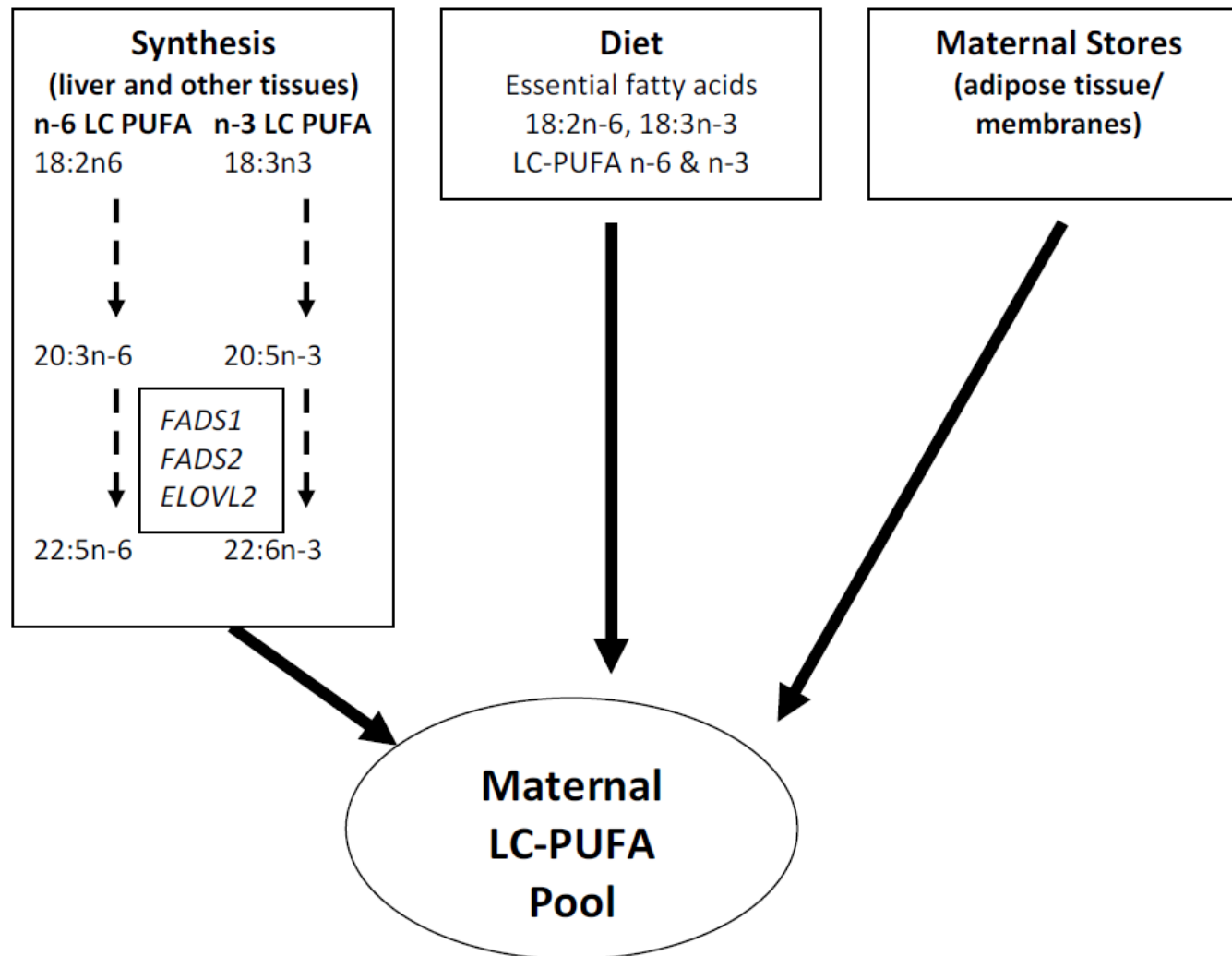
18:2n-6 (LA) ↑ 12% T1 – T3

18:3n-3 (ALA) ↑ 68% T1 – T3

26% increase in erythrocyte DHA concentration from end of first to the end of the third trimester

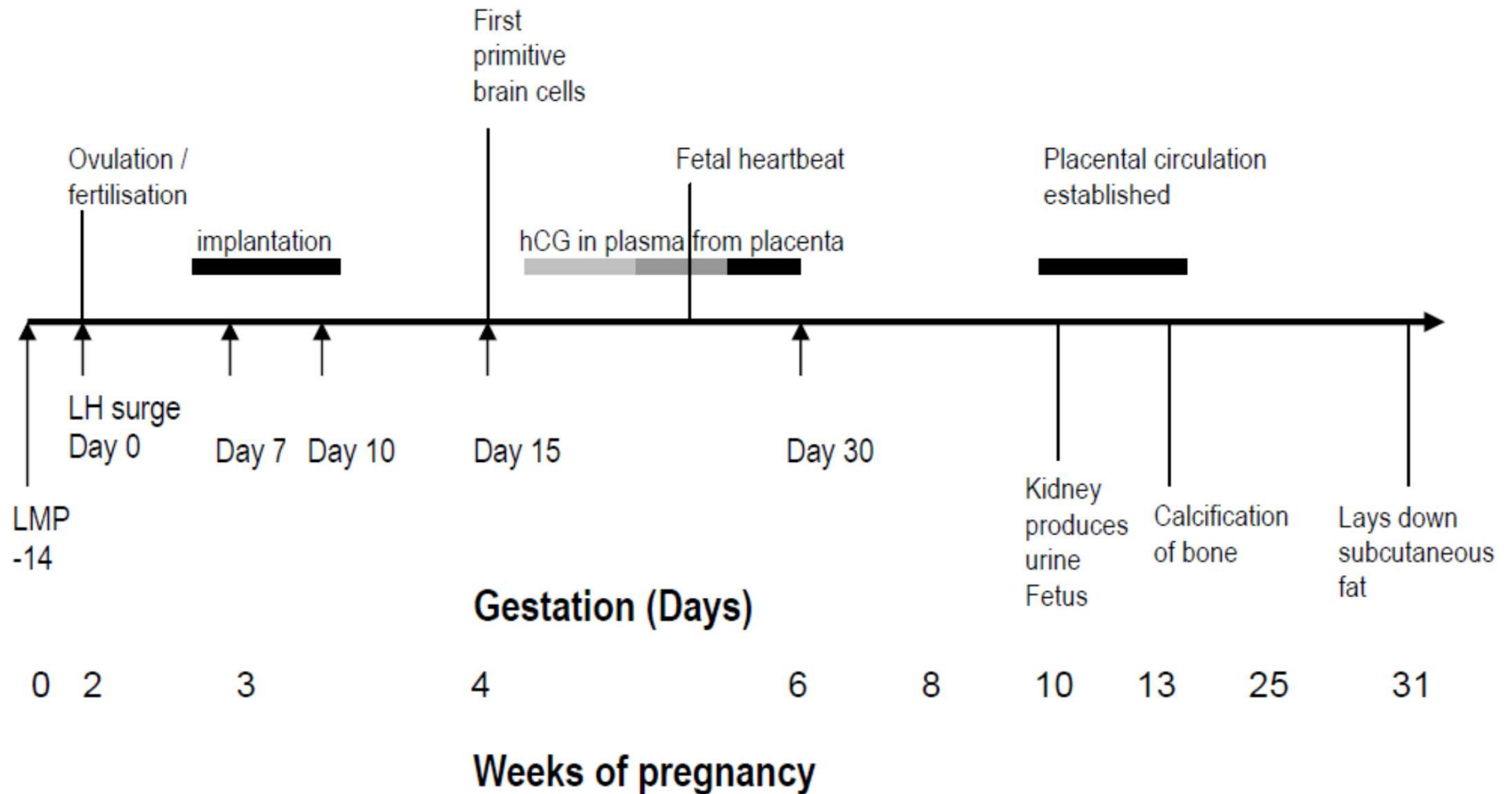
First trimester DHA is 17% above post natal levels

Sources of maternal DHA





Timeline of pregnancy





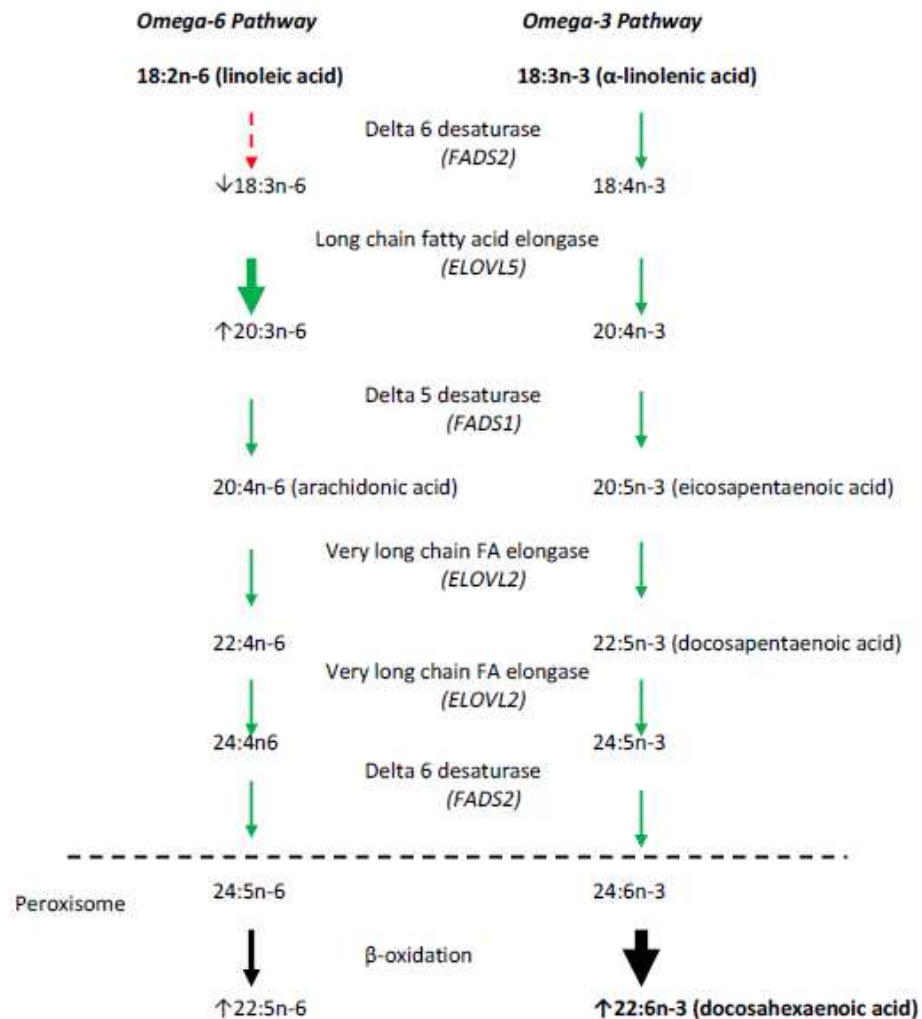
Rates of change of DHA concentration correlates with delta 6 desaturase activity

In rats, plasma and liver DHA levels and *FADS2* expression increased over gestation. *FADS2* expression correlated with oestradiol and progesterone.

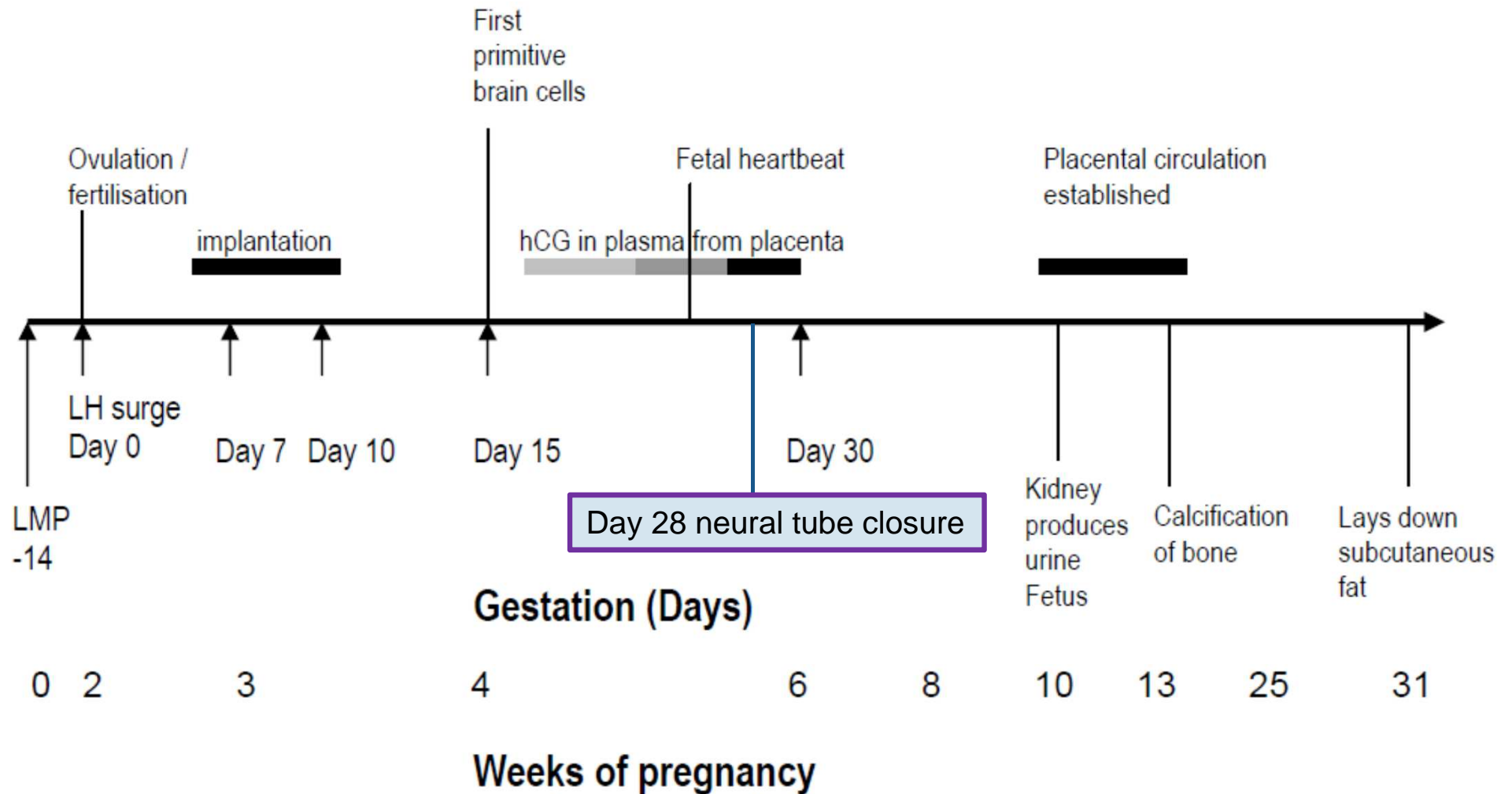
LA concentrations are decreased, relieving inhibition on n-3 pathway?

Polyunsaturated Fatty Acid Synthesis Pathways

(Mammalian)

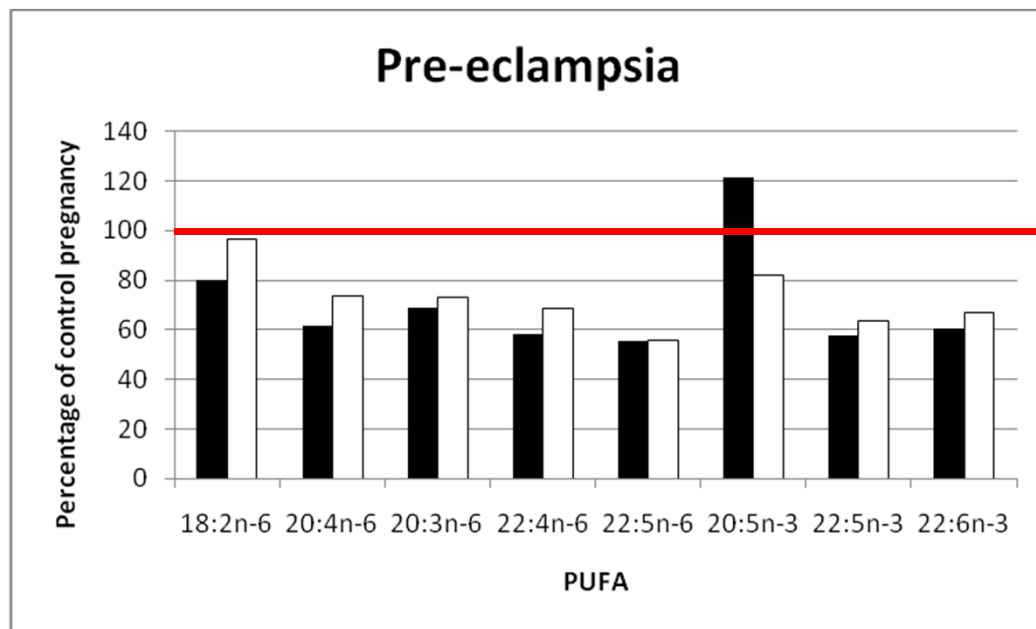


Timeline of pregnancy

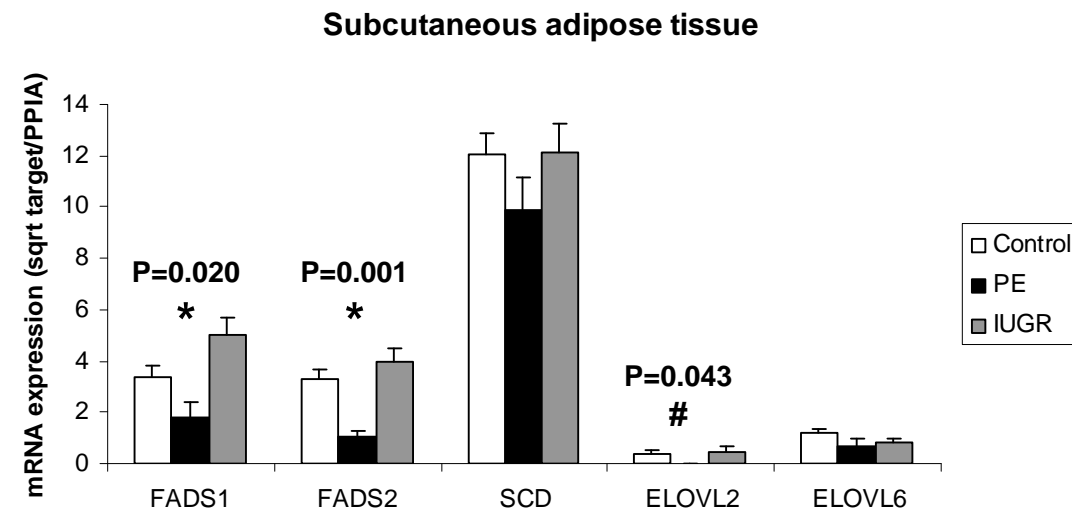


Reduced maternal and cord blood DHA in preeclampsia

Maternal and cord blood erythrocyte LC PUFA concentrations



Subcutaneous adipose tissue enzyme mRNA expression



FADS1 - $\Delta 5$ desaturase

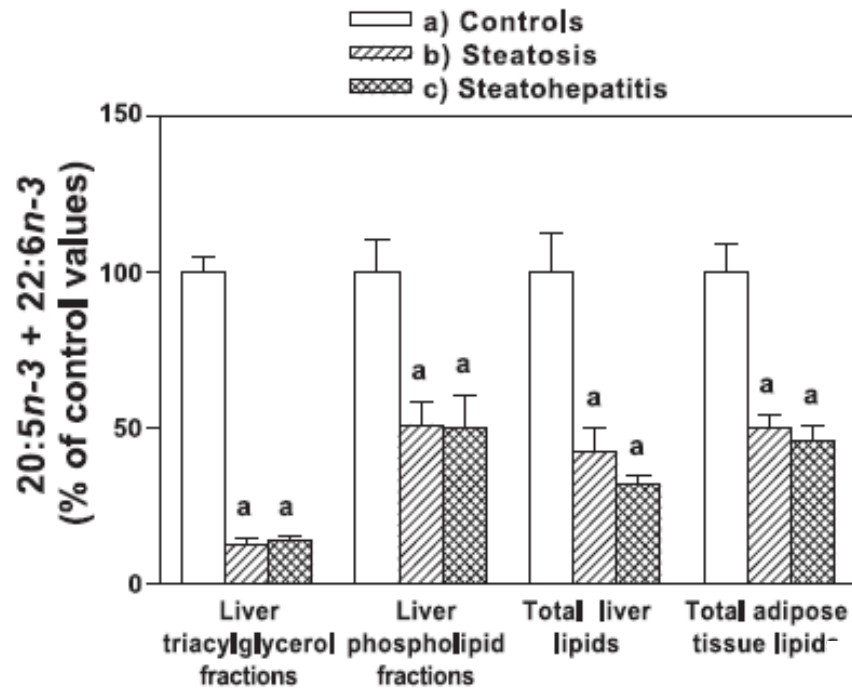
FADS2 - $\Delta 6$ desaturase

SCD - stearoyl coA desaturase

ELOVL2 - very long chain FA elongase

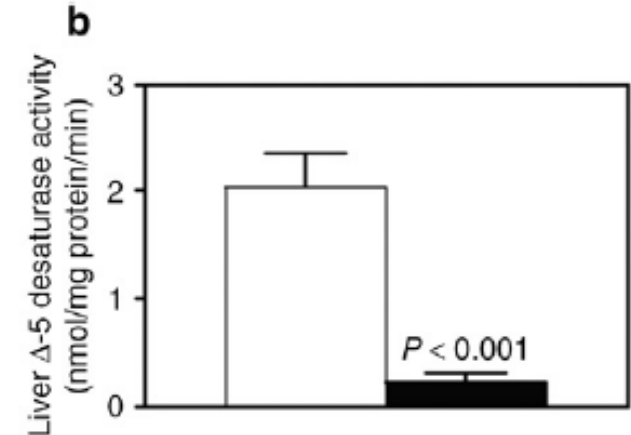
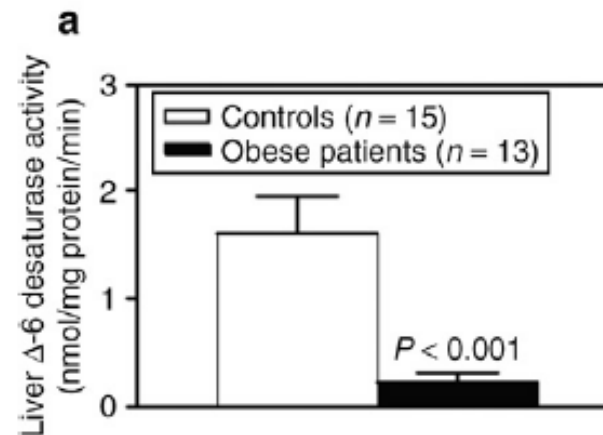
ELOVL6 - long chain FA elongase

Decreased synthesis of LC PUFA in non-alcoholic fatty liver disease (NAFLD)

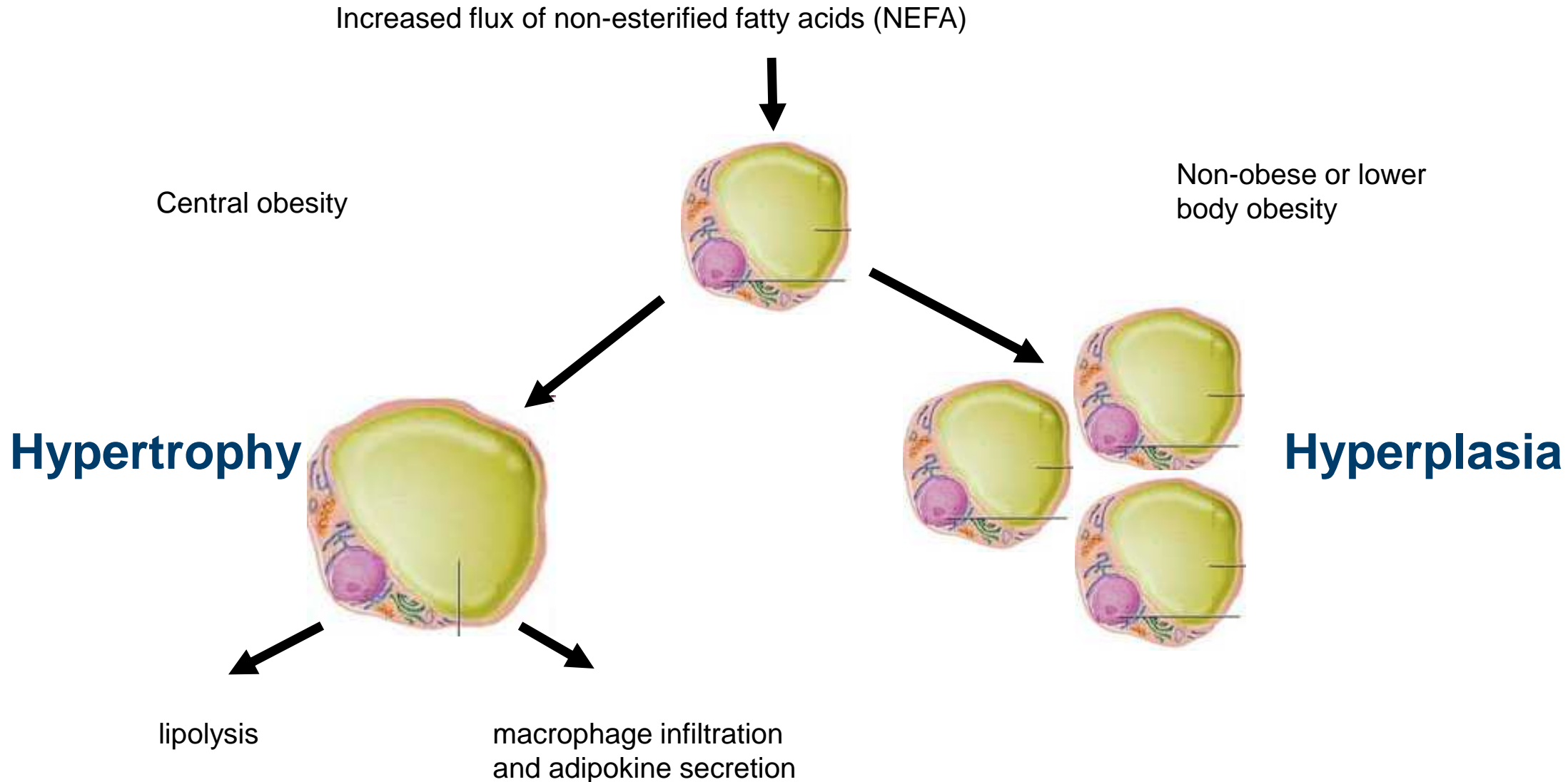


Videla et al, Free Radic Biol Med 2004

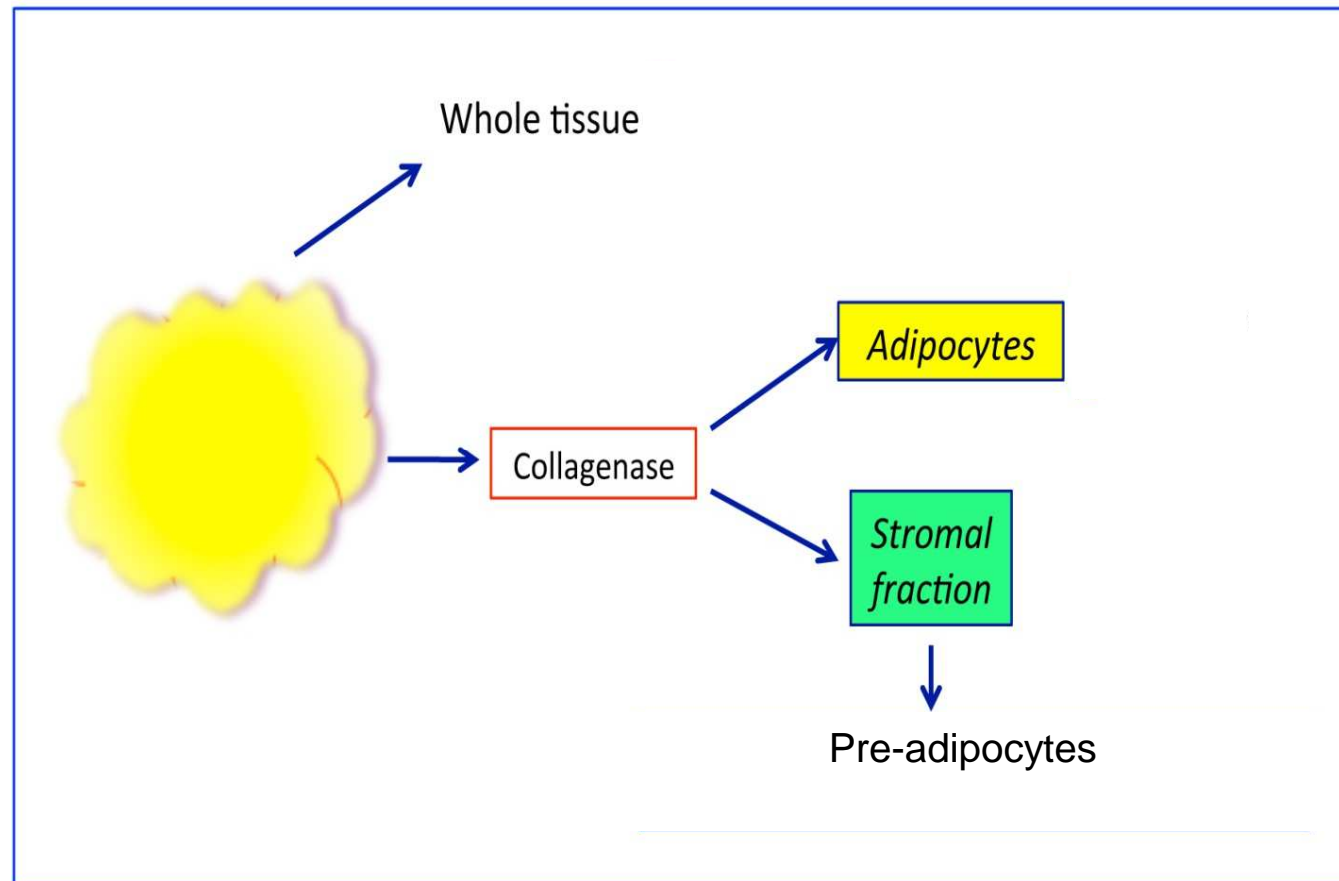
Araya et al, Obesity 2009



Hypertrophy vs hyperplasia of adipocytes



Adipose tissue is more than mature adipocytes



In the stromal fraction there are pre-adipocytes and macrophages

Adipocyte expandability

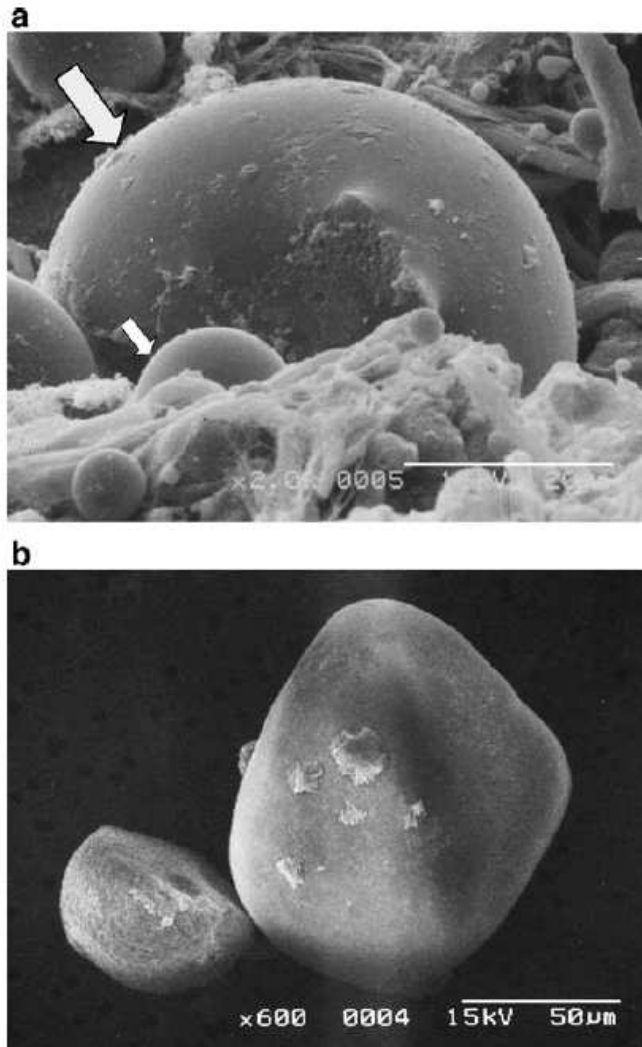


Fig. 2 Photographic examples of human adipose cells of widely varying cell size. **a** Scanning electron micrograph of paraformaldehyde-fixed tissue, showing small and very small adipose cells (arrows indicate cells of approximately 45 and 10 μm diameters). **b** Scanning electron micrograph of osmium-fixed cells, showing large and small adipose cells

- Insulin resistant individuals have more small adipocytes and decreased expression of genes related to adipose cell differentiation
- Individuals who are unable to recruit an additional population of mature adipocytes for TG storage are hypothesised to develop insulin resistance.

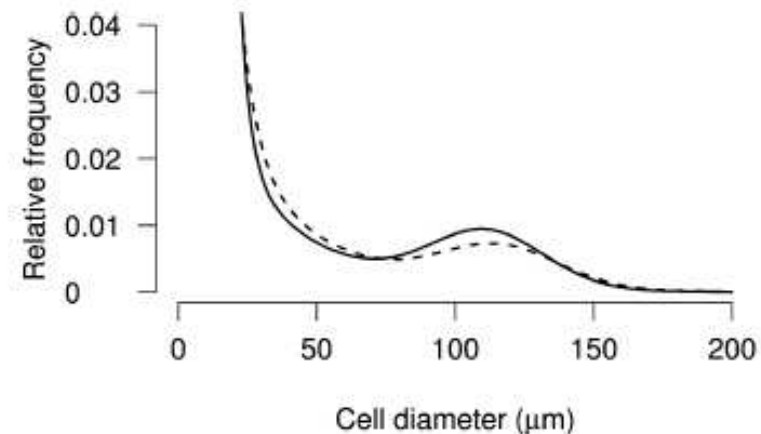


Fig. 4 Multisizer profiles of the adipose cell-size using the mean parameters from the curve-fitting formula for insulin-sensitive (*solid line*) and insulin-resistant (*dashed line*) subjects ($p=0.03$ using MANOVA)

The ability of fat depots to store fat in the subcutaneous depot



Healthy weight



Obese with adipocyte expansion

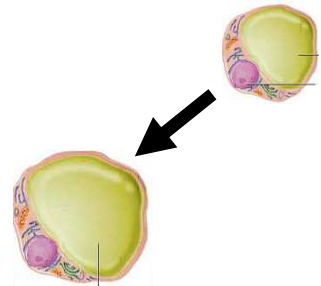


Obese with failure to differentiate
preadipocytes

Is adipocyte function the link between NAFLD and preeclampsia?

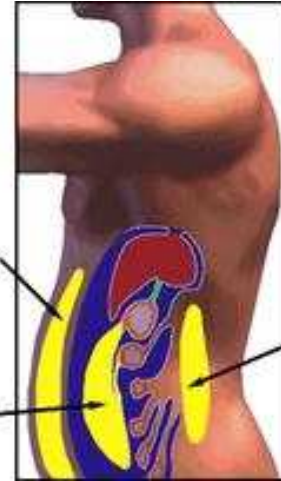
Gestational fat deposition

Hypertrophic obesity

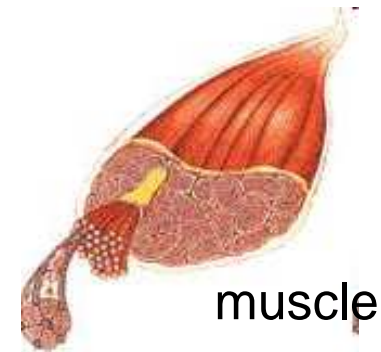
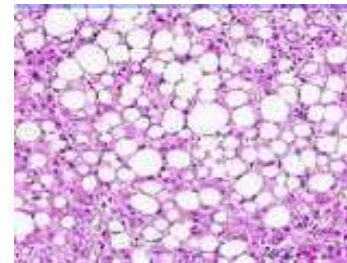


Subcutaneous

Visceral



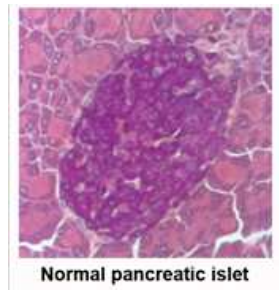
Ectopic lipid accumulation
also in placenta



muscle

NEFA spillover

pancreatic beta cell



More small adipocytes in preeclampsia - reduced ability to expand?

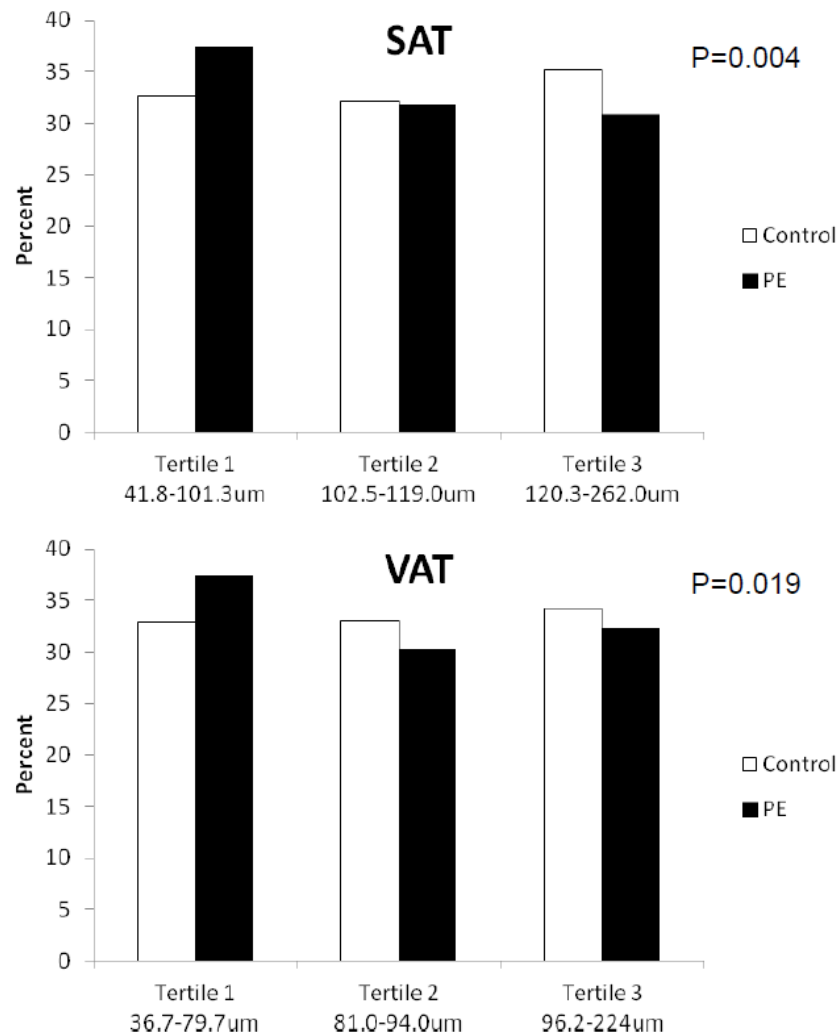
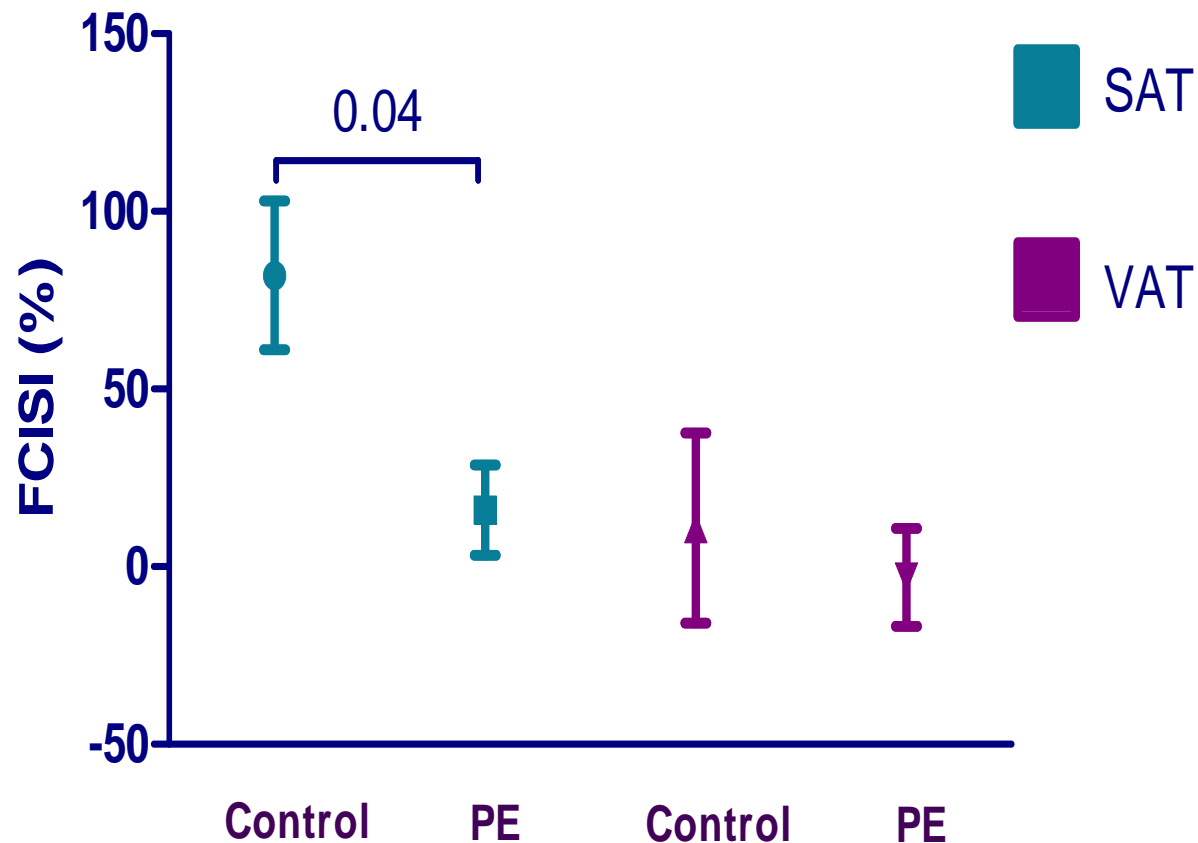


Figure S1. Subcutaneous adipose tissue (SAT) and visceral adipose tissue (VAT) adipocyte diameter distribution in control and preeclamptic (PE) pregnancy. Adipocyte diameter was measured in n=100 adipocytes from each adipocyte preparation. SAT and VAT adipocyte diameters were divided into tertiles and percent adipocytes within the diameter ranges calculated for healthy and PE samples. Percentage adipocytes in each tertile for the whole control and PE groups are shown.

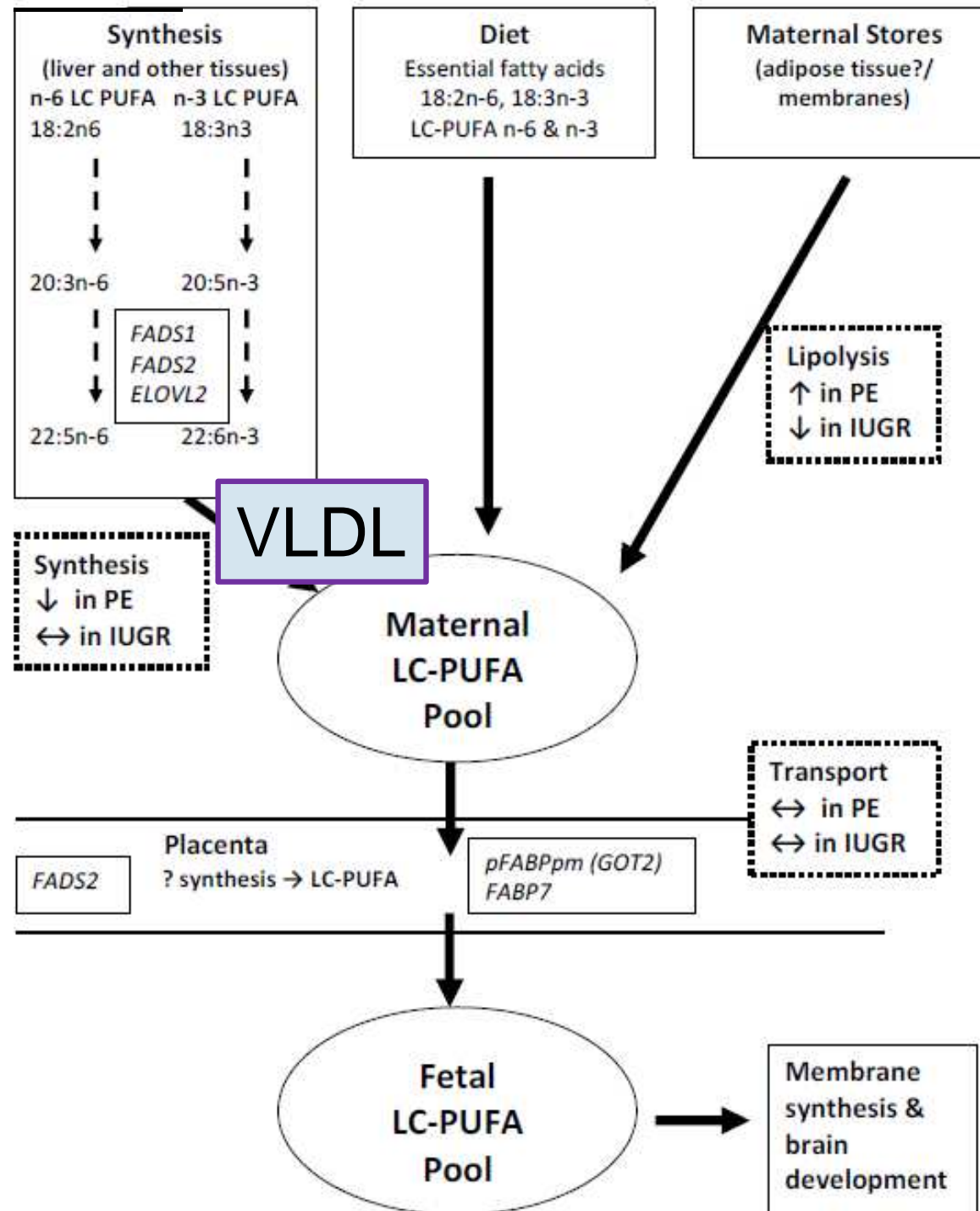
Adipocyte insulin resistance in PE

FCISI
Control vs. PE

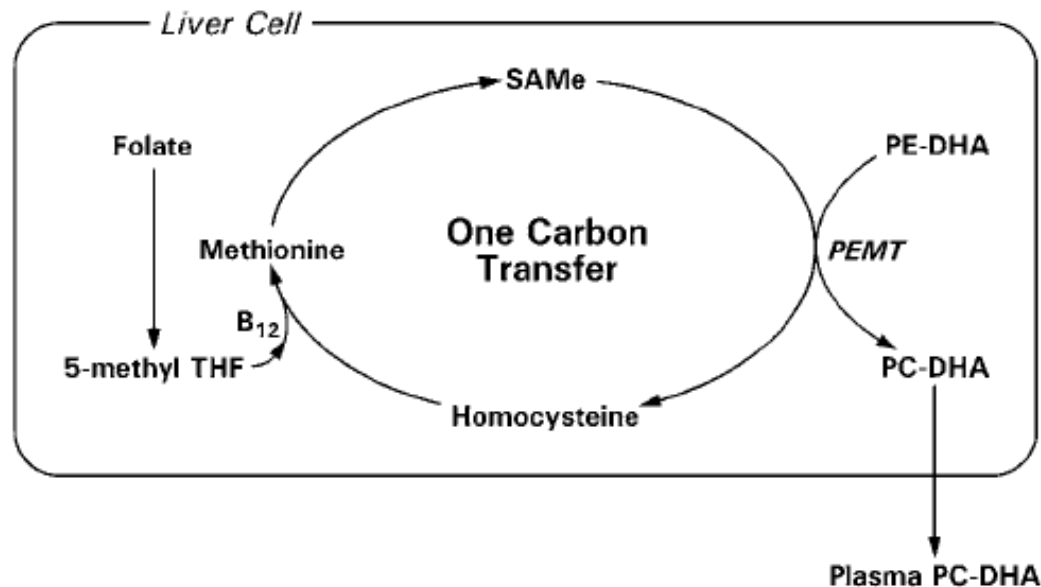
Fat cell insulin sensitivity index (FCISI) –
the ability of insulin to suppress beta
adrenergic stimulated lipolysis



In PE, SAT adipocytes as insulin
resistant as VAT



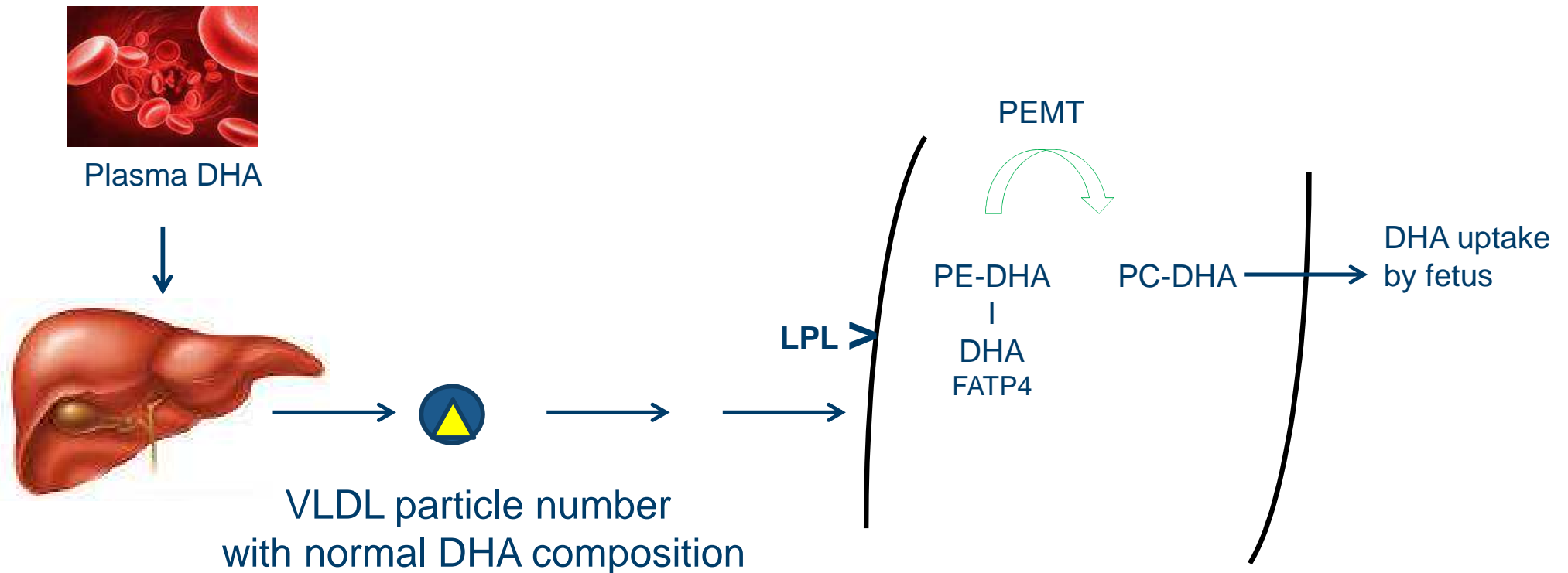
DHA mobilisation from the liver

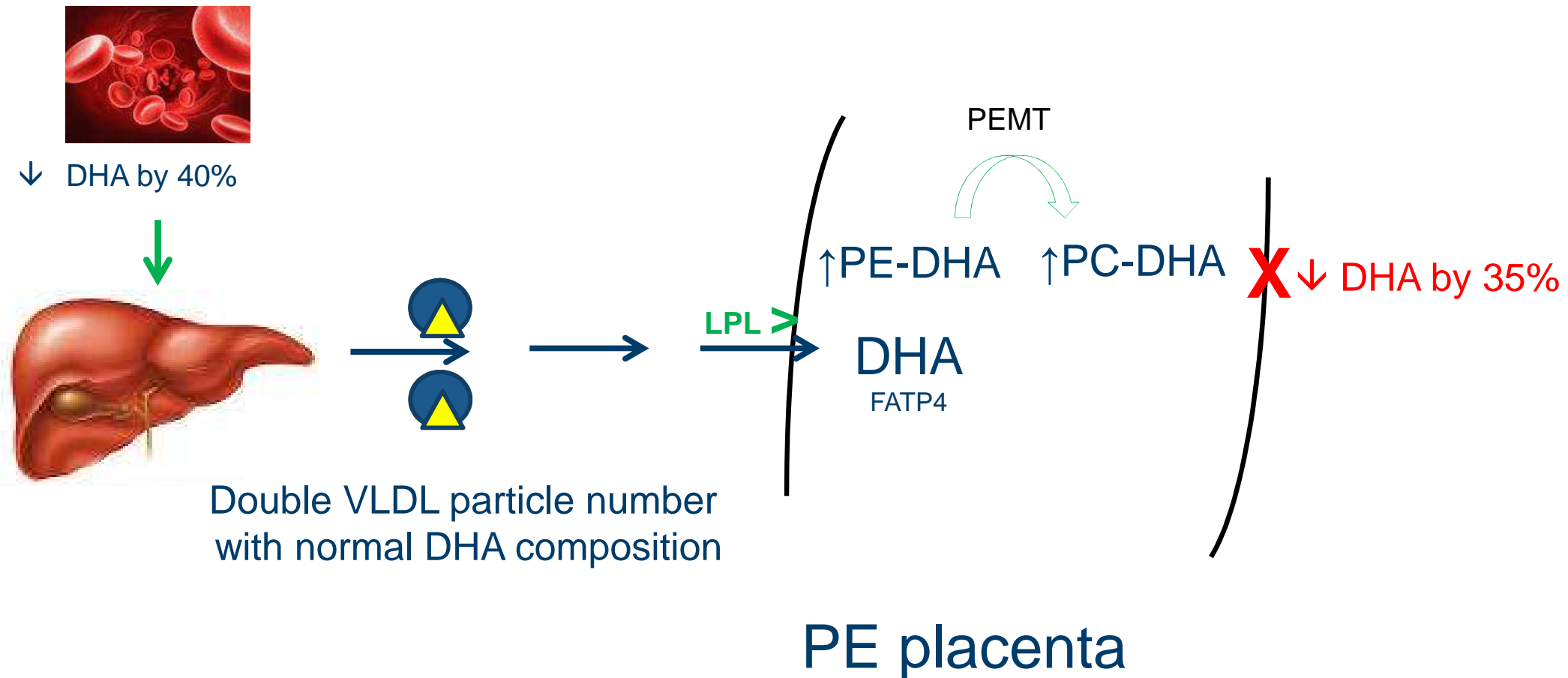


Folate and phosphatidylethanolamine-*N*-methyl transferase (PEMT) are involved in mobilising DHA from the liver into the plasma

Figure 2 Methyl transfer in the liver producing phosphatidylcholine-DHA is critical for mobilization of DHA into the blood. Dietary folate is converted in the body to 5-methyl tetrahydrofolate (5-methyl THF). Methyl transfer from 5-methyl THF to homocysteine requires vitamin B₁₂ and results in the synthesis of methionine. Methionine is converted into *S*-adenosyl-L-methionine (S-AdoMet). Methyl groups from S-AdoMet are transferred by phosphatidylethanolamine-*N*-methyltransferase (PEMT) to ethanolamine in a series of steps that convert it to choline and produce homocysteine. In this way, liver DHA incorporated into phosphatidylethanolamine is transformed into the nonpolar phosphatidylcholine-DHA. DHA which undergoes this process can be released from the liver into the plasma.

Could this system exist in placenta?





- DHA is an important nutrient for neurodevelopment
- Humans do not synthesise DHA efficiently
- Maternal DHA plasma concentrations are increased during pregnancy as early as day 18 gestation
- DHA is selectively transported across the placenta
- Maternal and fetal DHA levels are lower in preeclampsia possibly due to decreased maternal synthesis and a defect in placental transfer

Glasgow

Ann Brown

Shahzya Huda

Fiona Jordan

Vanessa Mackay

Christopher Onyiaodike

Frances Stewart

Helen Lyall

Heather Murray

Scott Nelson

Rob Nibbs

Naveed Sattar

University of Wollongong

Barbara Meyer

Simon Brown

Sam Eather

Todd Mitchell

