



2020 ACS Spring National Meeting & Expo, Philadelphia

[MEDI 268] Biochemical, biophysical and structural studies of hPRMT5 and its peptide–competitive MTA–synergistic small molecule inhibitors for cancer therapeutics

PRESENTER: Wei Zhou (Ph.D. candidate)

DIVISION: Division of Medicinal Chemistry

SESSION: General Orals (MEDI)

DATE: Wednesday, March 25, 2020

University of Florida

College of Medicine

Department of Biochemistry and Molecular Biology

Chenglong Li Lab

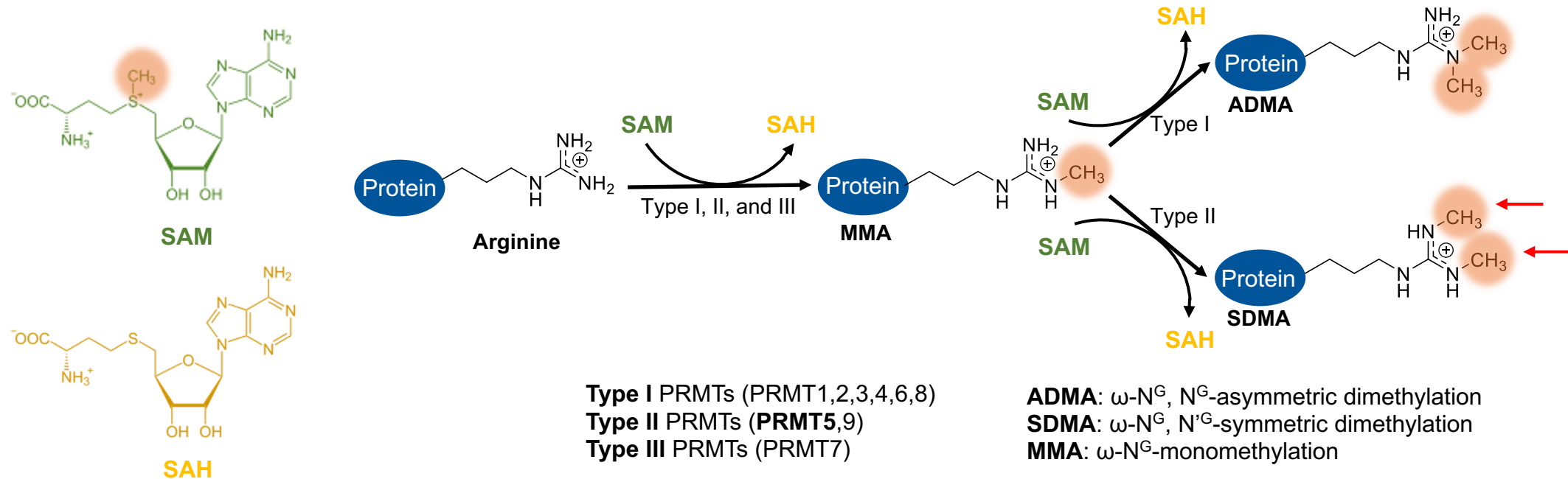
OUTLINE

- **Background**
- **Aim**
- **Methods and Results**
- **Conclusions**
- **Future Direction**

Arginine methylation by PRMT5

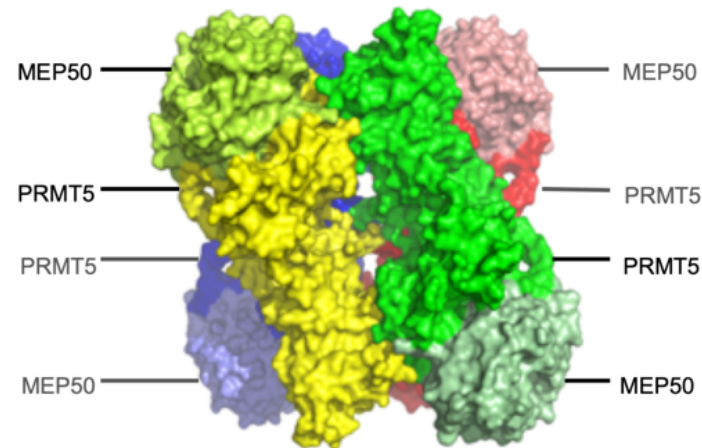
PRMT5 (Protein arginine methyltransferase 5)

- The main symmetric dimethylarginine (**SDMA**) methyltransferase (Type II)
- S-Adenosyl-L-Methionine (**SAM**) ==> S-Adenosyl-L-Homocysteine (**SAH**)
- Major substrates: H4R3, H2AR3, H3R2, H3R8 and non-histone proteins, e.g. p53.

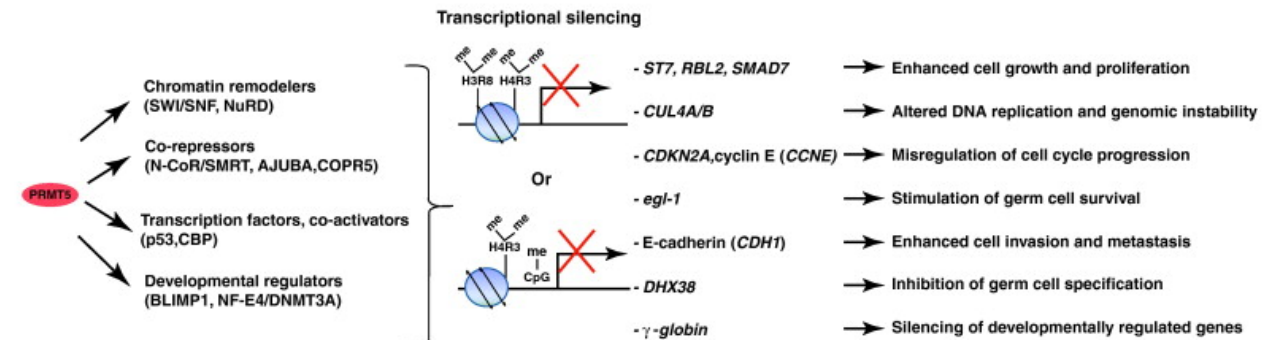
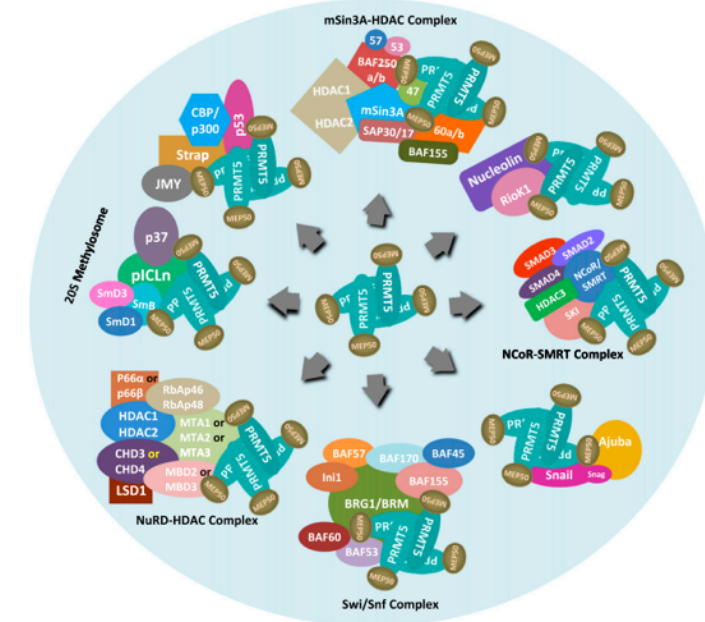


PRMT5 in cancers

- PRMT5 forms a hetero-octameric complex with MEP50.
- PRMT5:MEP50 complex forms large multimolecular complexes with different binding partners.

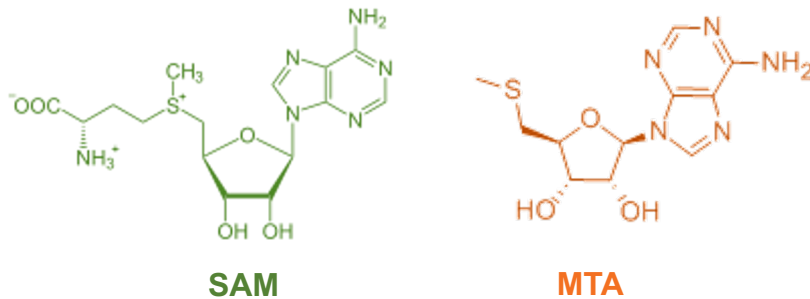


- PRMT5 is overexpressed in many cancers.
- PRMT5 inhibition or depletion induces cell growth arrest and cell death.



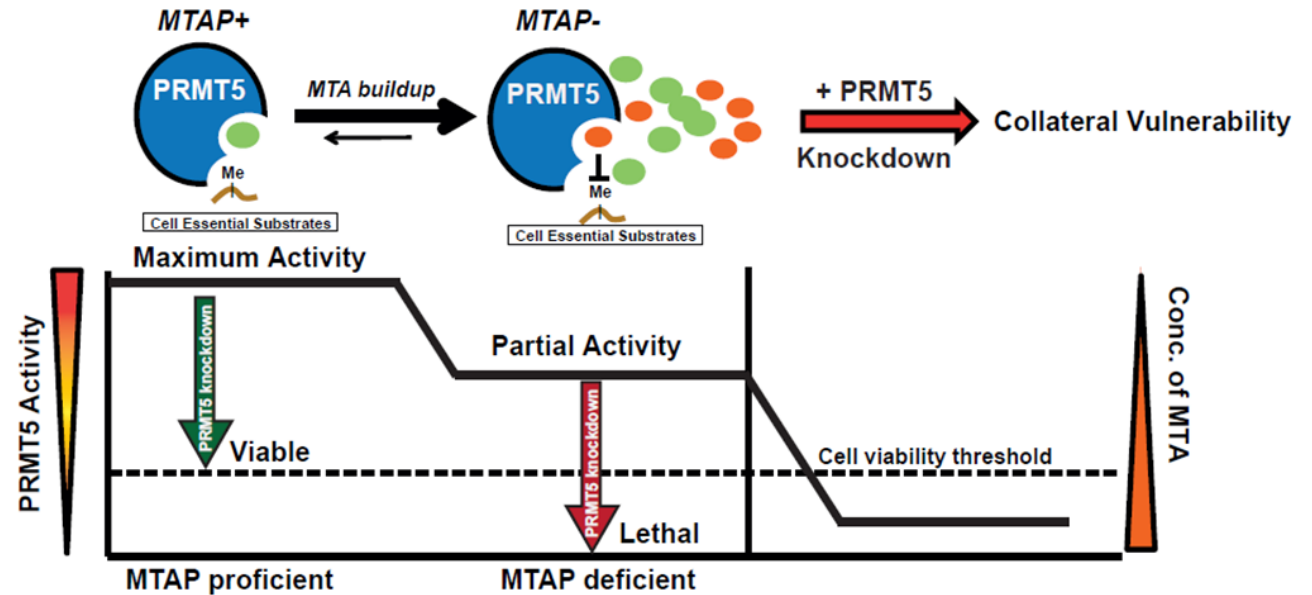
PRMT5 as a novel drug target in MTAP-deleted cancers

- Methylthioadenosine phosphorylase (MTAP) deletion occurs in ~15% of all human cancers.
- MTAP-deletion in cancer cells disorders methionine metabolism and results in **MTA accumulation** in cells.
- MTA creates a **partially inhibited enzyme state** thus significantly increases their vulnerability to further PRMT5 inhibition.



Intracellular concentration:

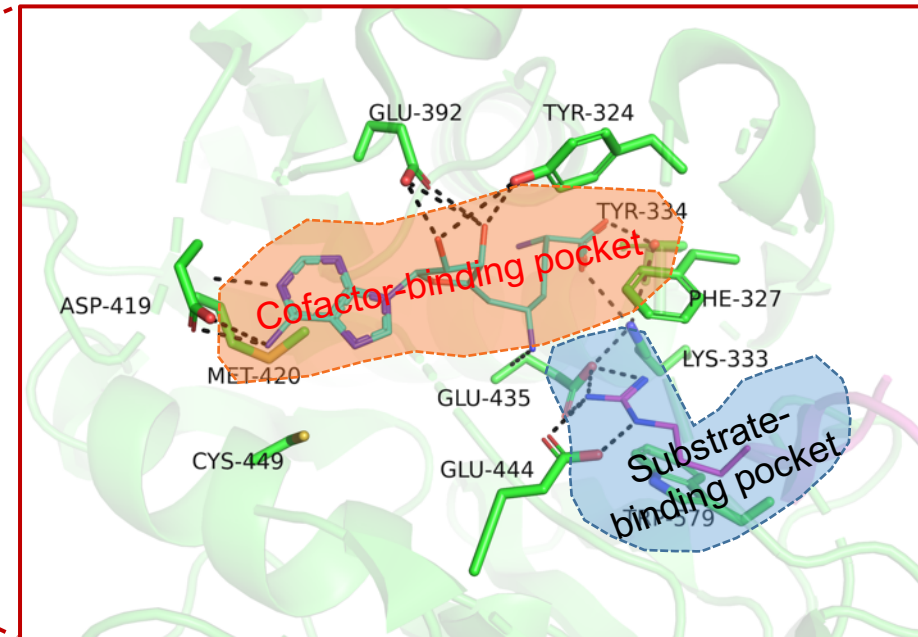
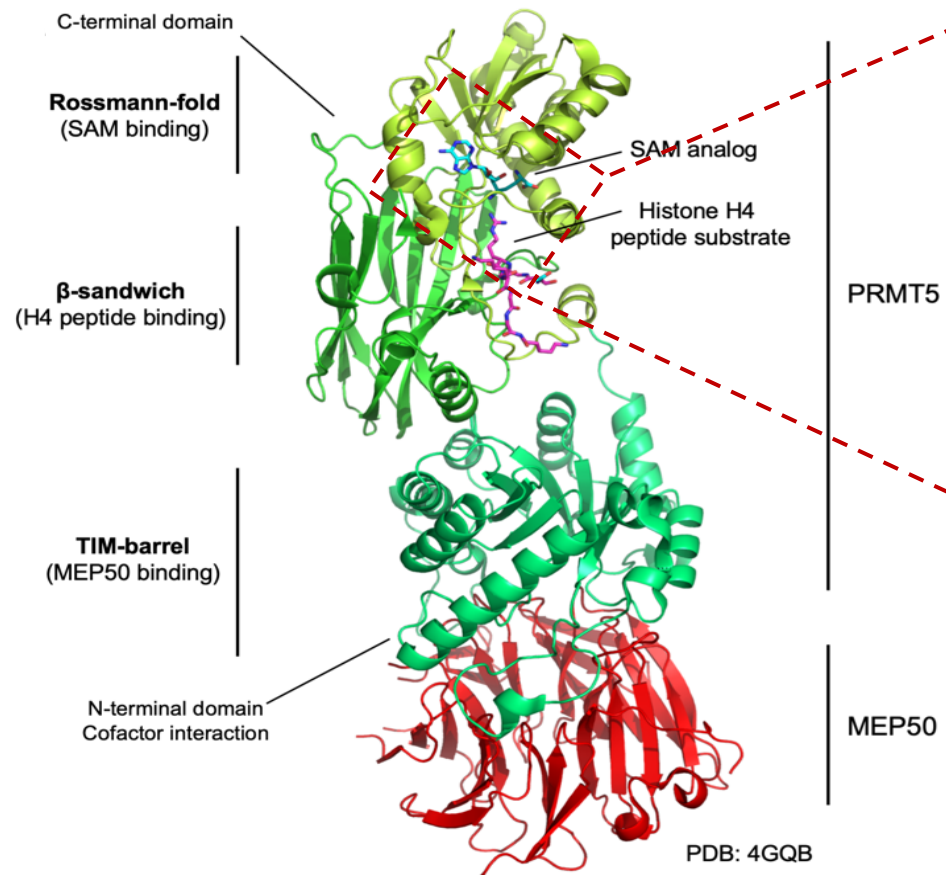
	SAM	MTA
MTAP w.t.	~50 μ M	~1 μ M
MTAP -/-	~50 μ M	~20 μ M



As an oncoprotein in tumorigenesis, PRMT5 emerges as a compelling target for drug discovery in cancer therapy.

Crystal structure of PRMT5:MEP50

The first crystal structure of human PRMT5:MEP50 was published in 2012.

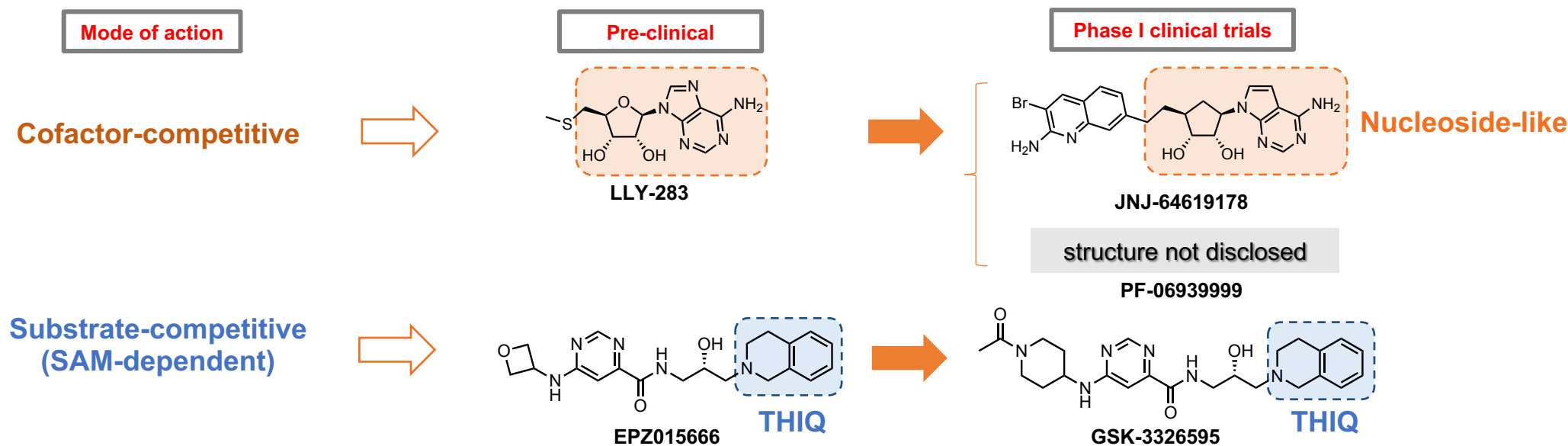
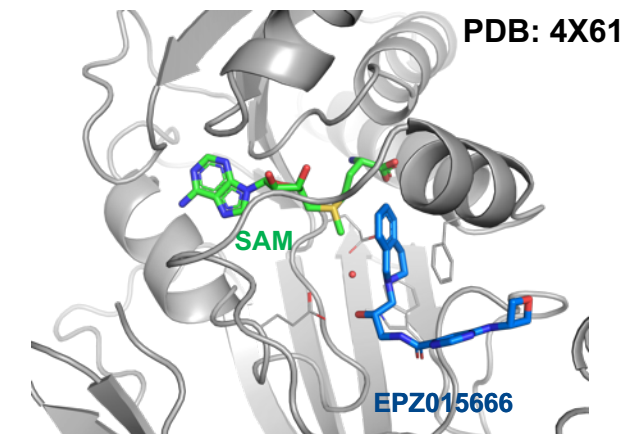


- Crystal structure of human PRMT5:MEP50 in complex with a cofactor analog (cyan) and a H4 substrate peptide (magenta).
- PRMT5 (green) consists of a TIM-barrel domain for MEP50 (red) binding, a Rossmann-fold domain for cofactor binding and a β -sandwich domain for substrate peptide binding.

Limitations of PRMT5 inhibitors in clinical trial

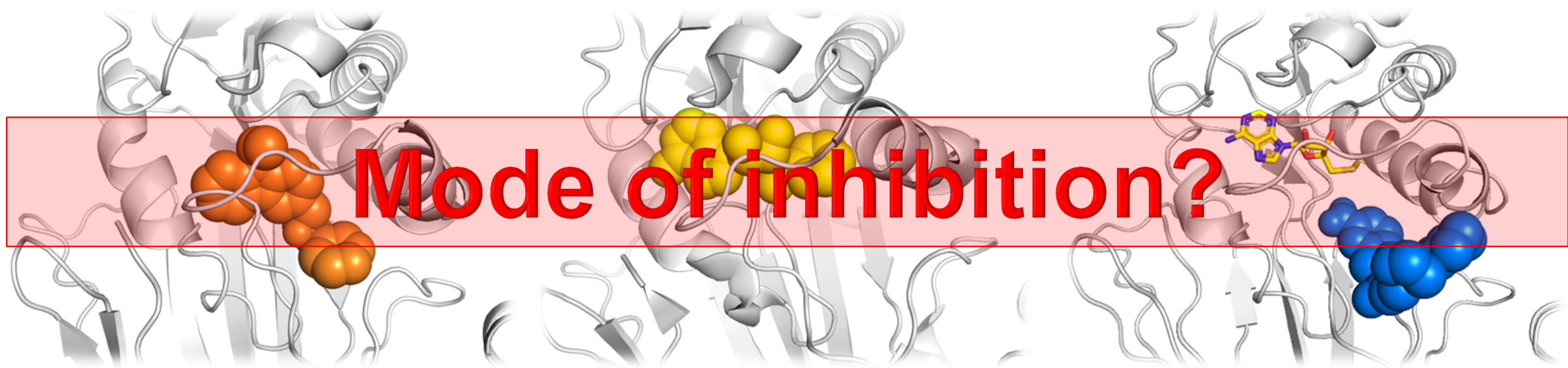
Since 2016, three drug candidates have entered **Phase I clinical trial**:

- **Cofactor-competitive inhibitors** (JNJ-64619178 and PF-06939999)
SAM-analogs may cause off-target effect.
- **Substrate-competitive inhibitors** (GSK-3326595)
Pre-binding of cofactor SAM is required to form cation- π interactions;
Low binding affinity to MTA-bound form (MTAP-deleted cancers).



AIM

Design a potent, non-nucleoside, SAM-independent small molecule inhibitor targeting PRMT5.



Identification of CMP5 as a 'hit'

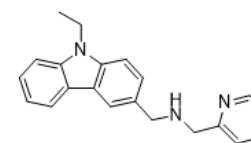
Identification of hit compound CMP5 by virtual screening and cellular assays

Virtual screening the ChemBridge CNS-Set library of 10,000 small molecule compounds at PRMT5 active site (cofactor+arginine)

Top compounds with lowest binding energy from the screen were visually inspected for contacts that mimic conserved PRMT5-SAM-ARG interactions

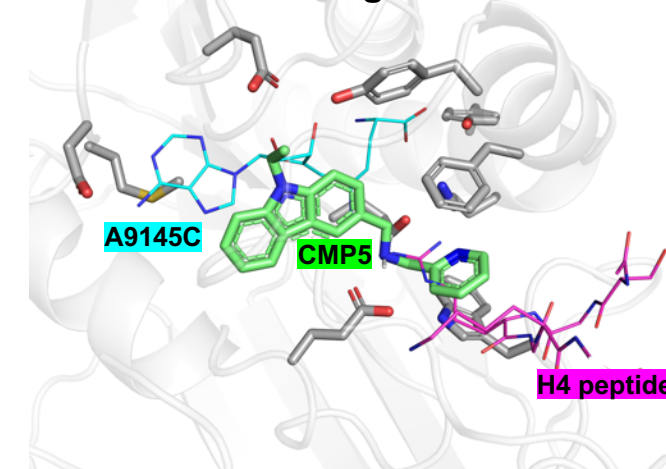
8 potential compounds were selected for cellular screening assays using immunofluorescence

Hit compound:
CMP5



CMP5

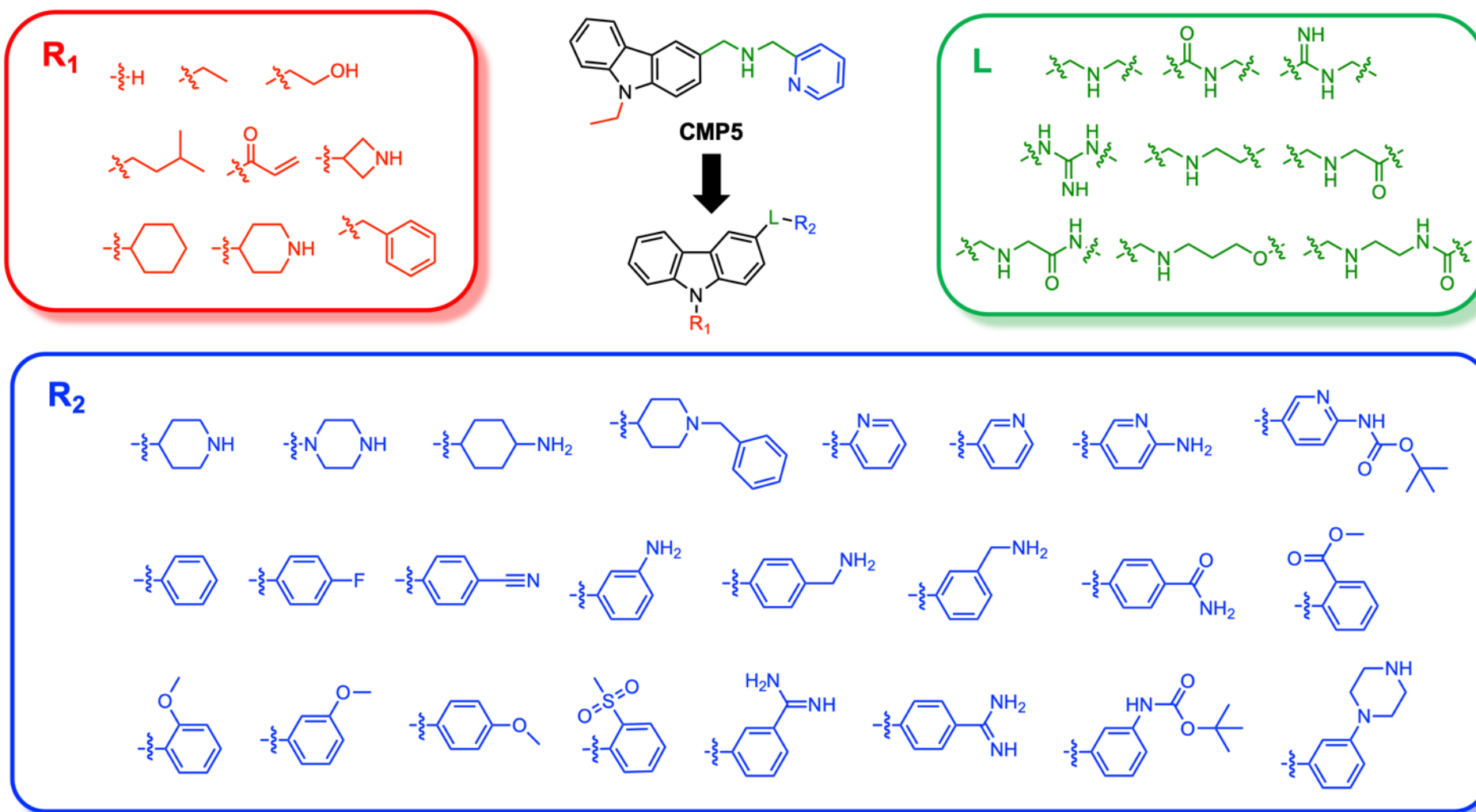
Predicted binding mode of CMP5



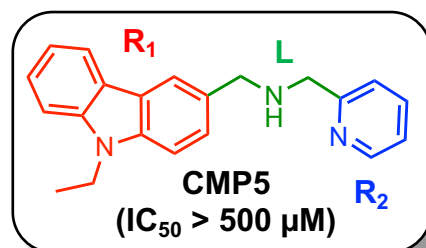
Cellular assays:

- ✓ Histone methyltransferase activity assay using hSWI/SNF-associated PRMT5;
- ✓ Selectivity against other Type I and Type II PRMTs;
- ✓ Cell proliferation assay;
- ✓ Apoptosis analysis;
- ✓ Immunofluorescence;
- ✓ Confocal microscopy;
- ✓ Western blot;
- ✓ Immunoprecipitation (IP).

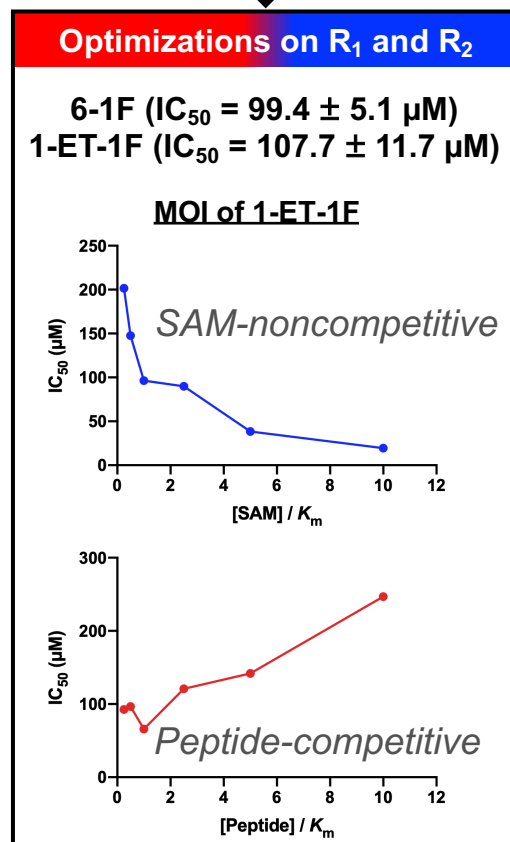
Structure-based drug design and structure-activity relationship study



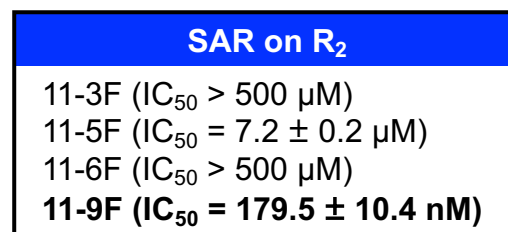
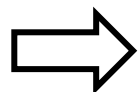
Compound optimization and mode of inhibition (MOI) study



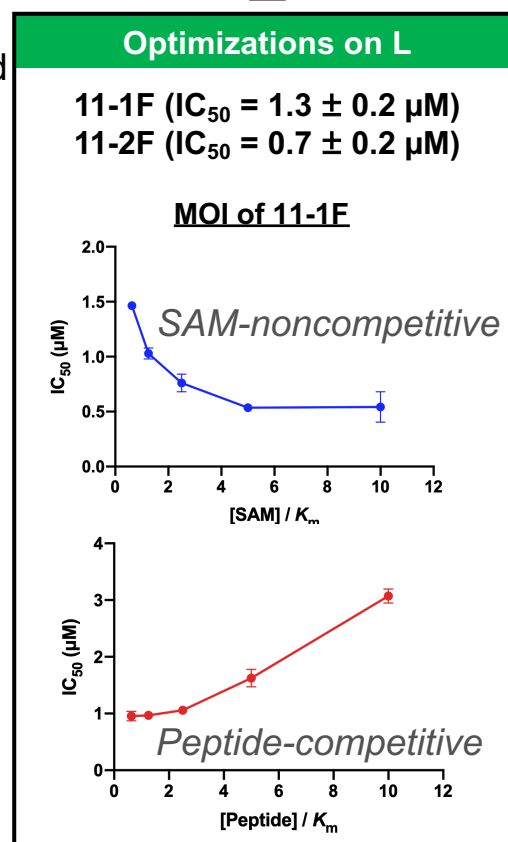
↓ > 5-fold



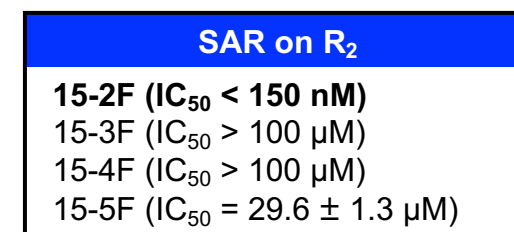
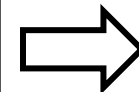
~100-fold



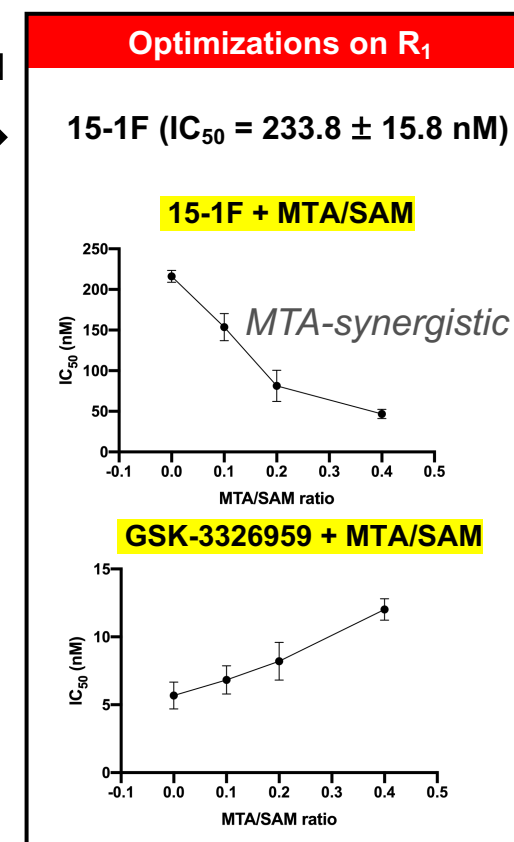
↑ ~5-fold



~5-fold



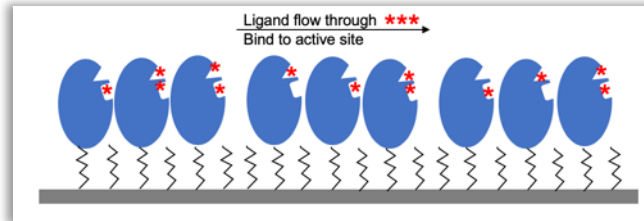
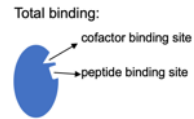
↑



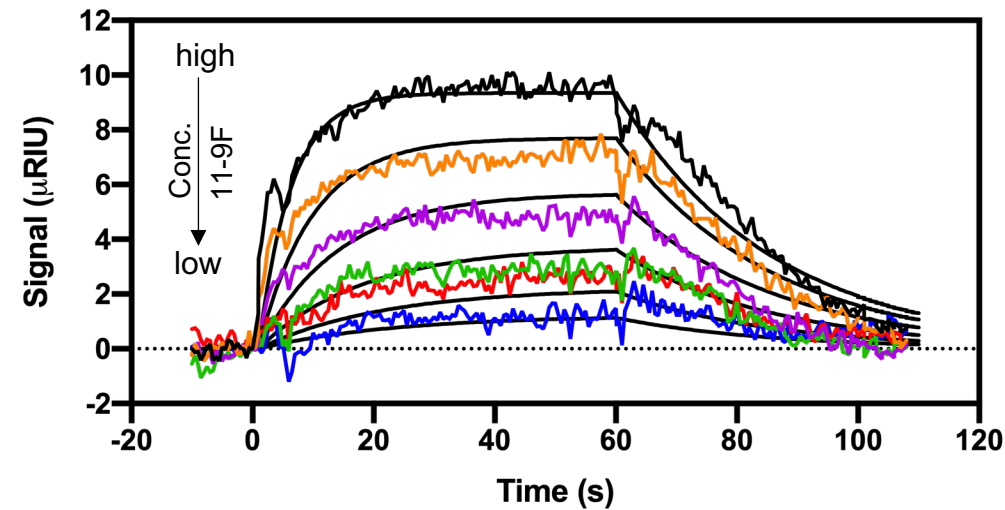
Binding preference study by SPR

Protein immobilized

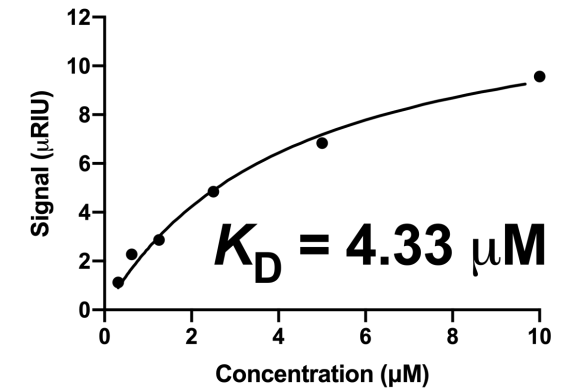
apo PRMT5:MEP50



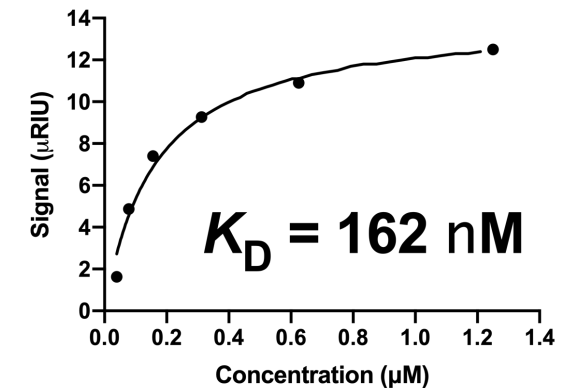
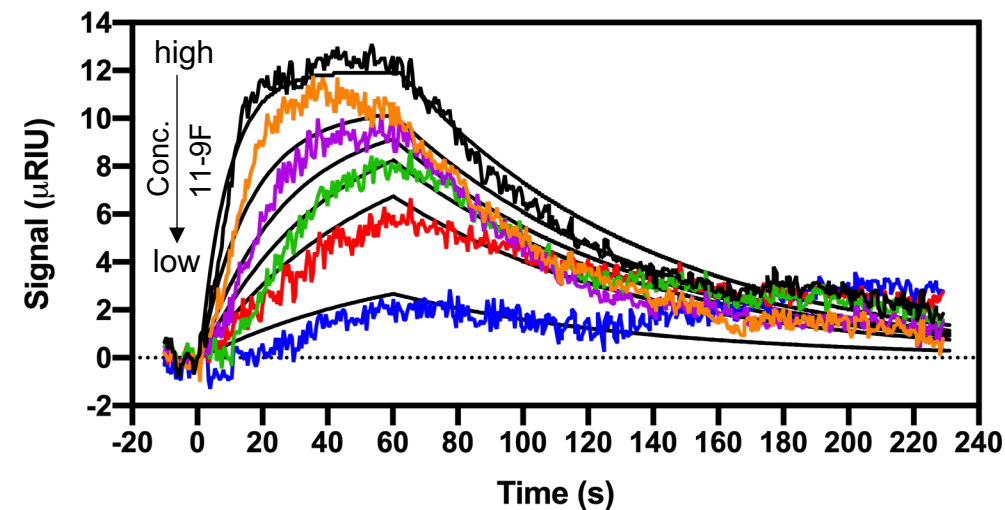
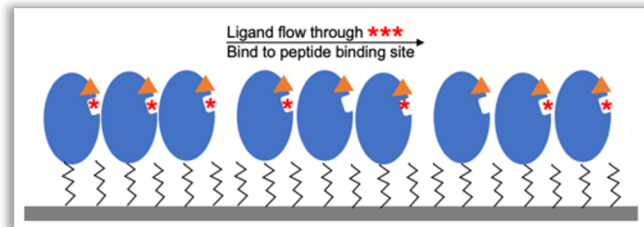
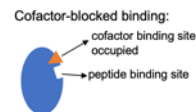
Sensorgram



K_D (affinity fitting)



PRMT5:MEP50
saturated with MTA



Binding preference study by SPR

Compound name	Enzymatic IC ₅₀ (μM)	Binding affinity K_D (μM) on different receptors			
		Apo P:M	P:M + MTA	P:M + SAH	P:M + SAM
11-1F	1.3	15.7	0.256	1.67	1.32
11-2F	0.7	12.1	0.095	0.895	0.692
11-3F	> 500	N.B	L.B	N.B	N.B
11-5F	7.2	N.B	N.B	N.B	2.84
11-6F	> 500	N.B	L.B	N.B	N.B
11-9F	0.180	4.33	0.162	0.191	0.732
15-1F	0.234	7.32	0.038	0.218	0.106
GSK-3326959	0.006	N.B	5.9	0.794	< 0.1

L.B.: very low binding response; N.B.: no binding

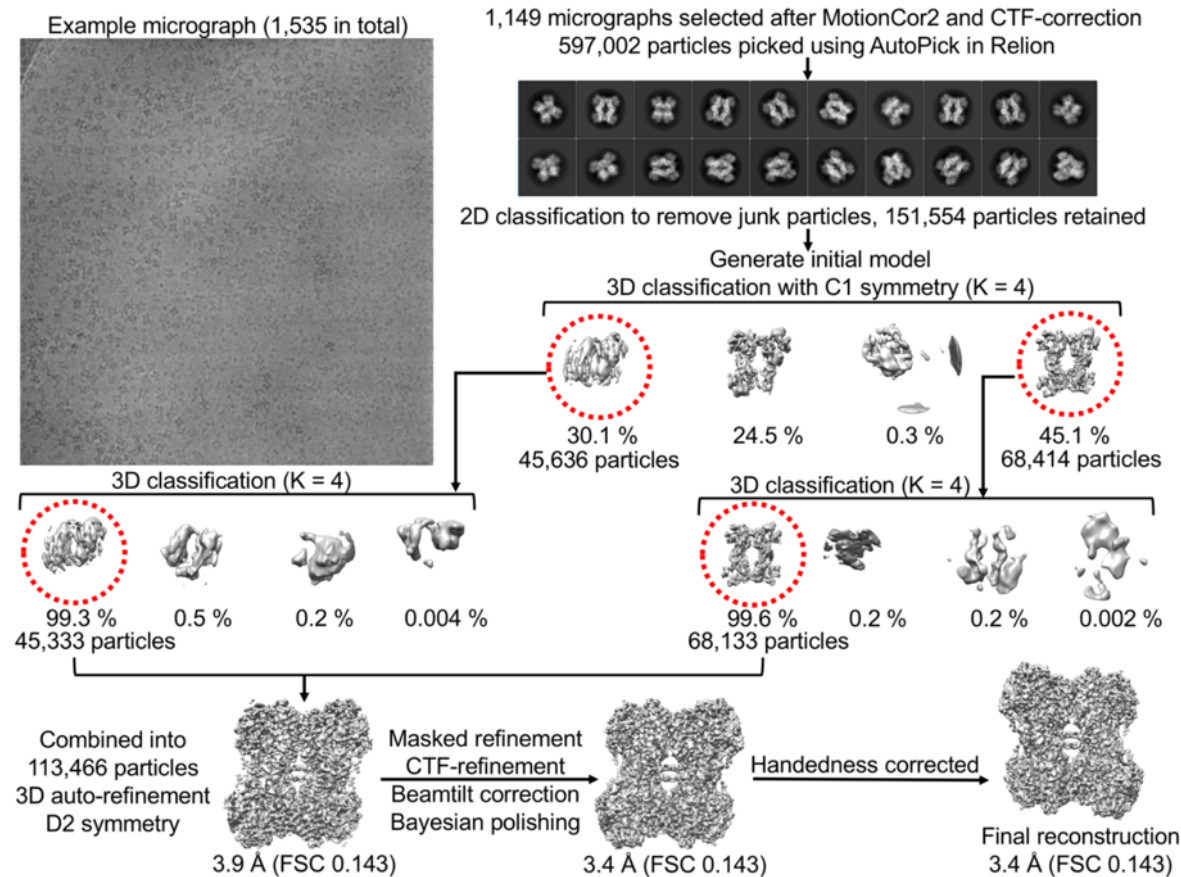
Binding preference study by SPR

Compound name	Enzymatic IC ₅₀ (μM)	Binding affinity K _D (μM) on different receptors				
		Apo P:M	P:M + MTA	P:M + SAH	P:M + SAM	
11-1F	1.3	15.7	0.256	1.67	1.32	
11-2F	0.7	12.1	0.095	0.895	0.692	
11-3F	> 500 (inactive)	N.B	L.B	N.B	N.B	No binding
11-5F	7.2	N.B	N.B	N.B	2.84	
11-6F	> 500 (inactive)	N.B	L.B	N.B	N.B	No binding
11-9F	0.180	4.33	0.162	0.191	0.732	
15-1F	0.234	7.32	0.038	0.218	0.106	
GSK-3326959	0.006	N.B	5.9	0.794	< 0.1	

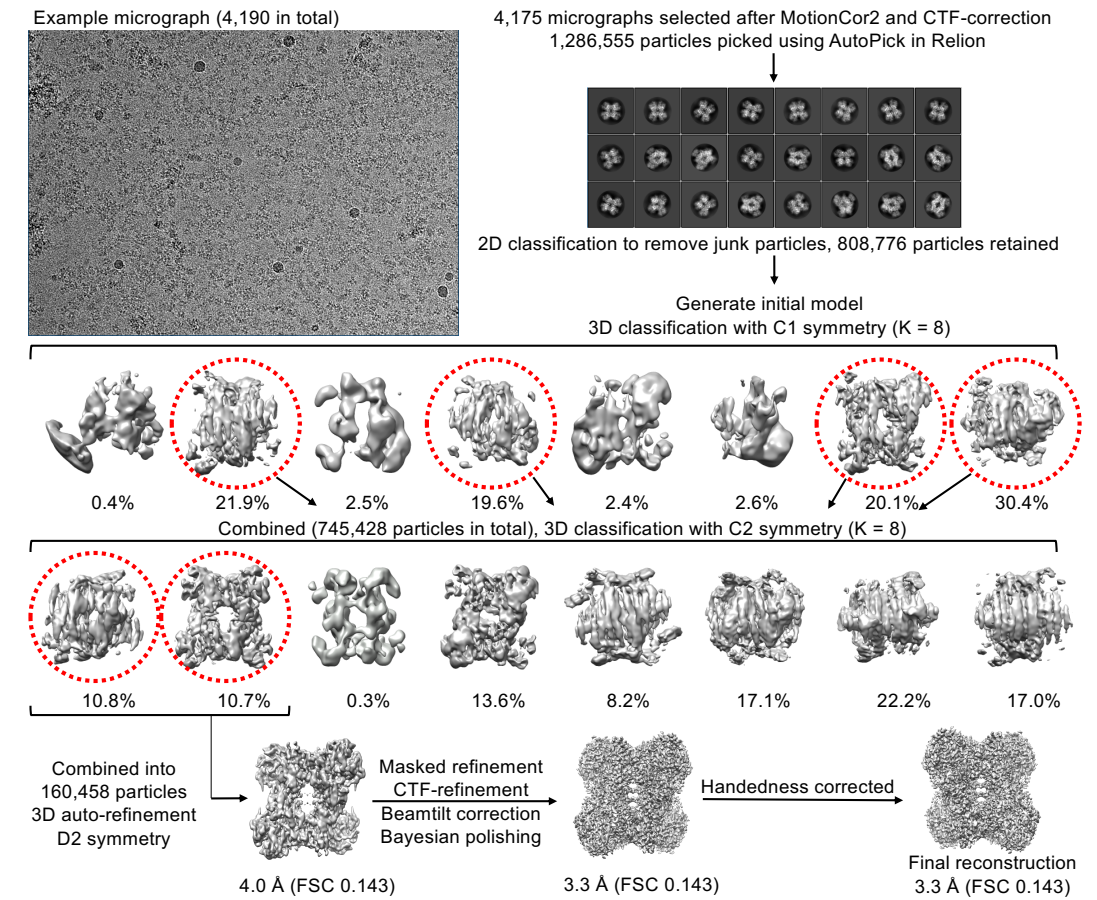
L.B.: very low binding response; N.B.: no binding

Structure determination by cryo-EM

The *apo* form of PRMT5:MEP50

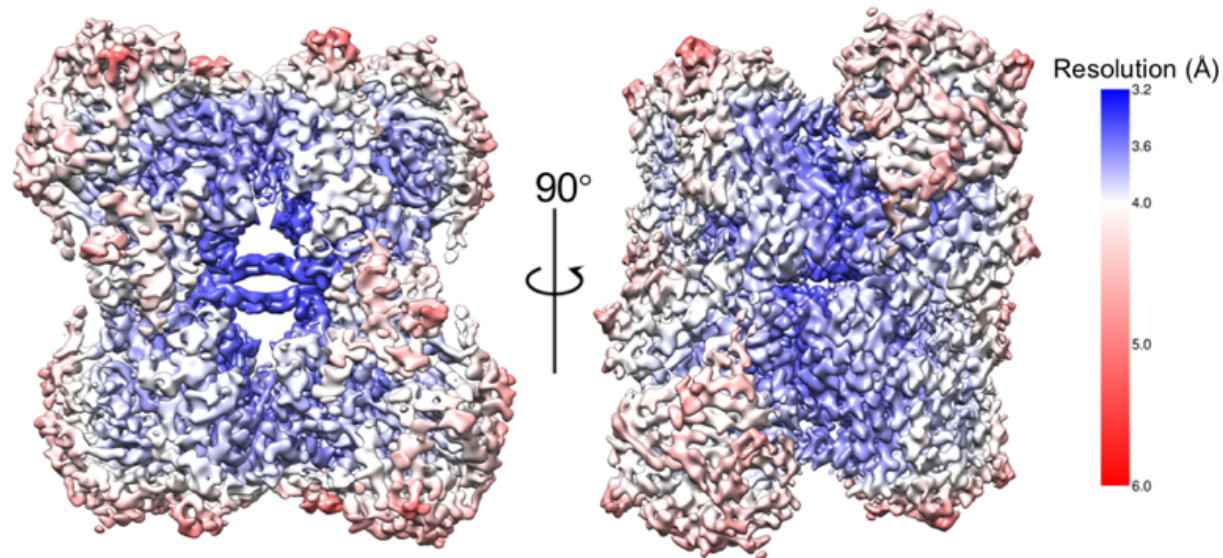


PRMT5:MEP50 in complex with MTA and 11-2F

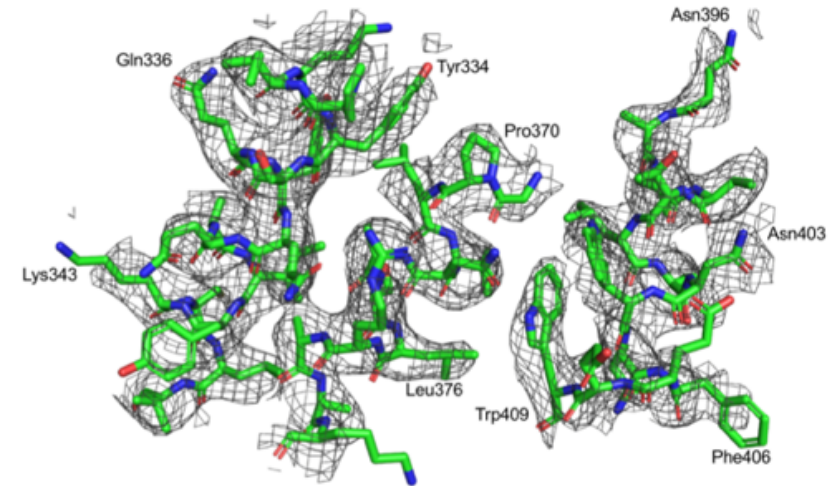


Structure determination by cryo-EM

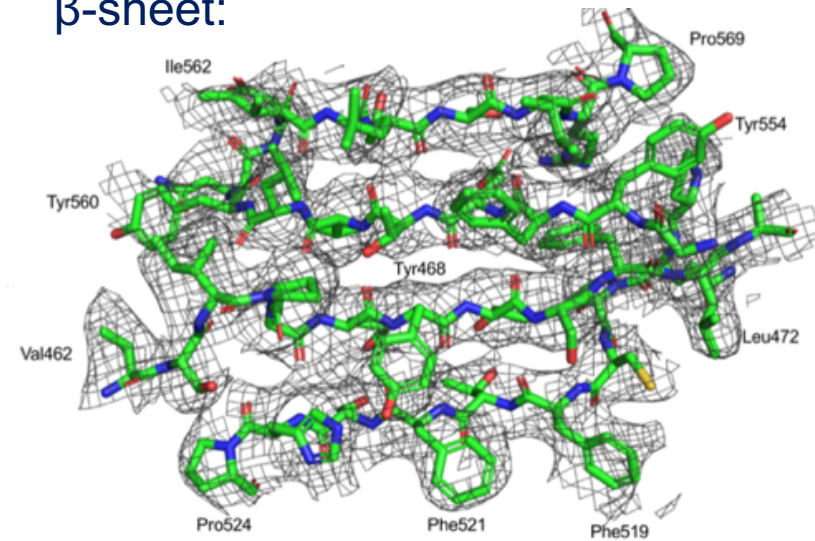
Local resolution of *apo* PRMT5:MEP50 complex:



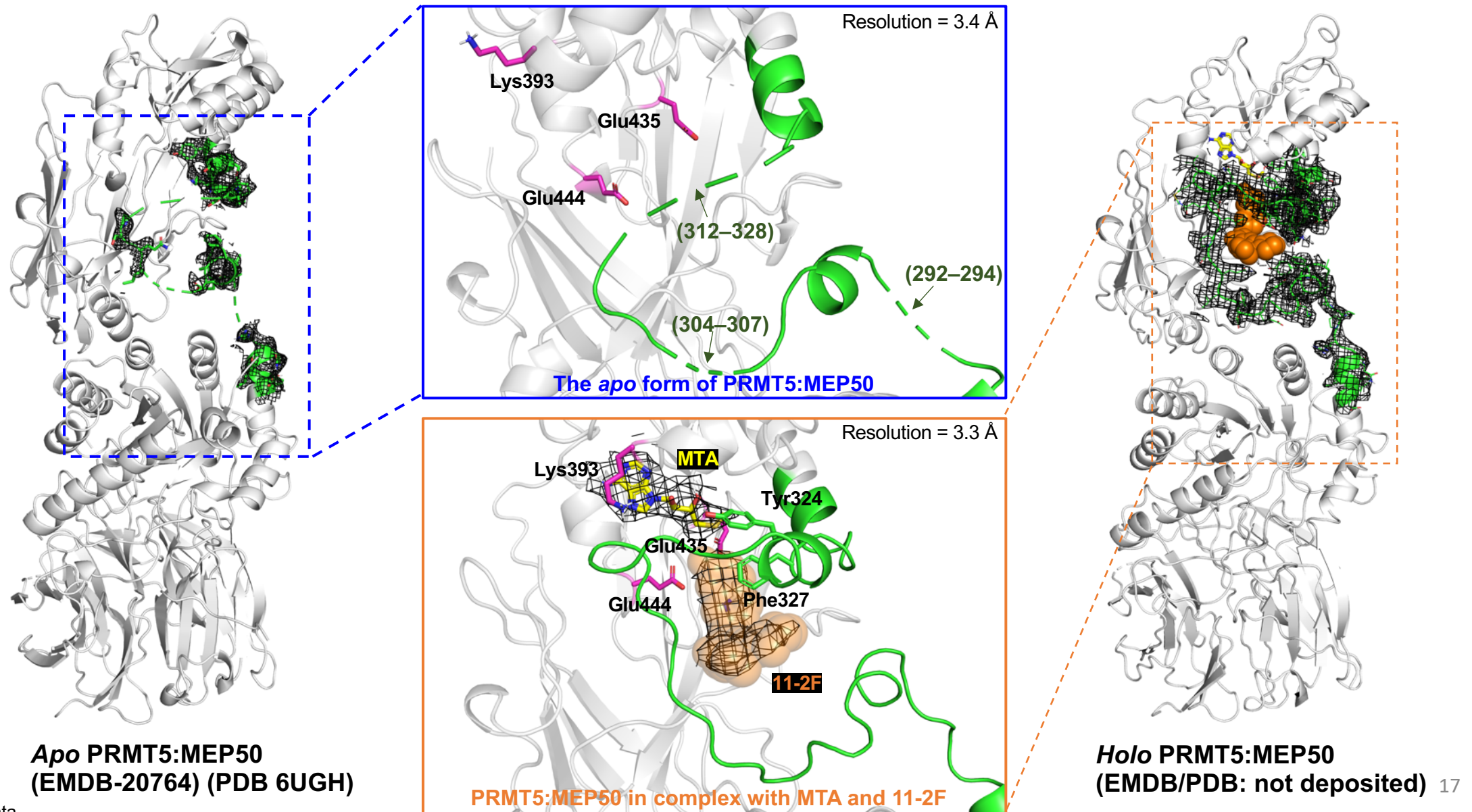
α -helix:



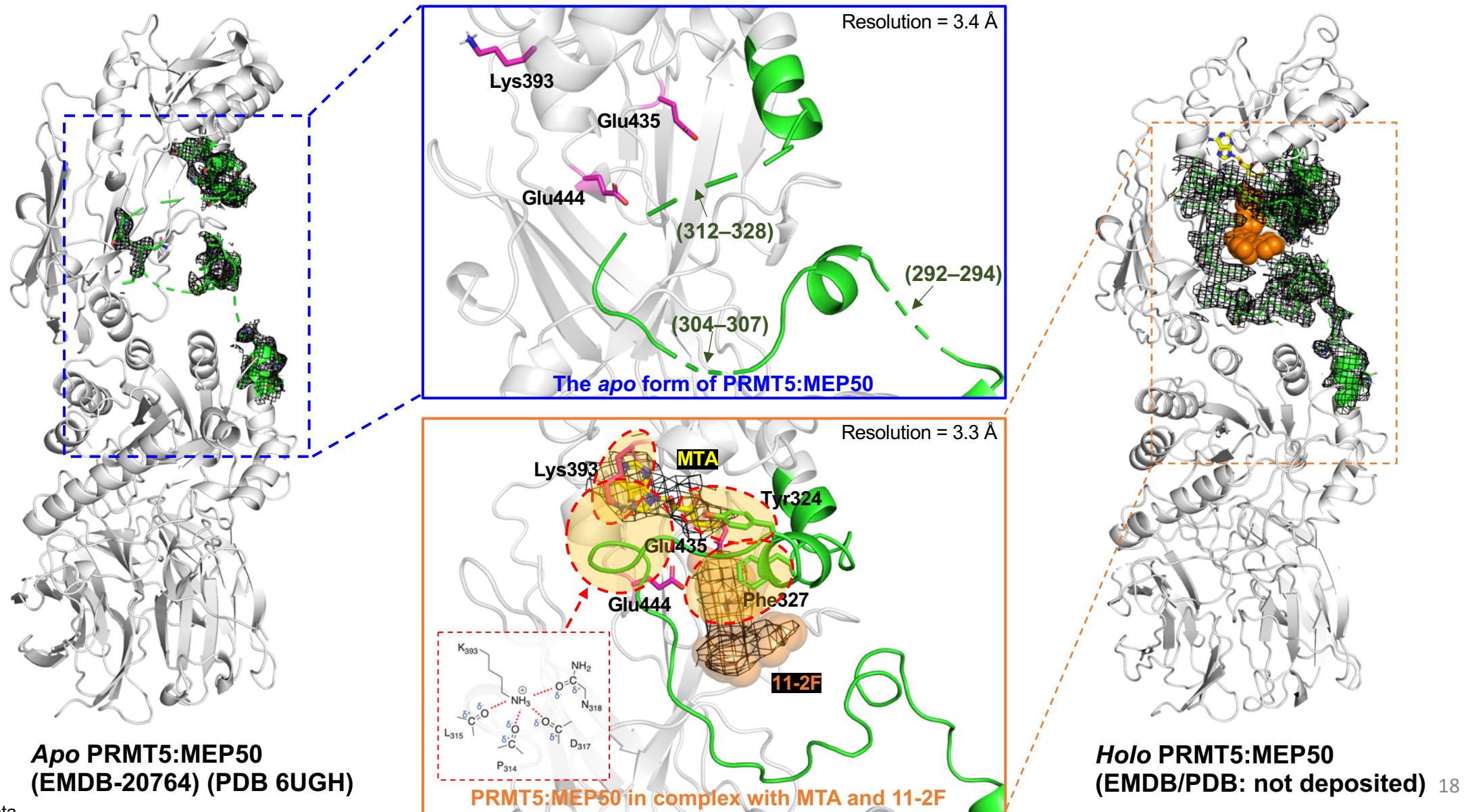
β -sheet:



Structure comparison of apo- and holo- forms of PRMT5:MEP50

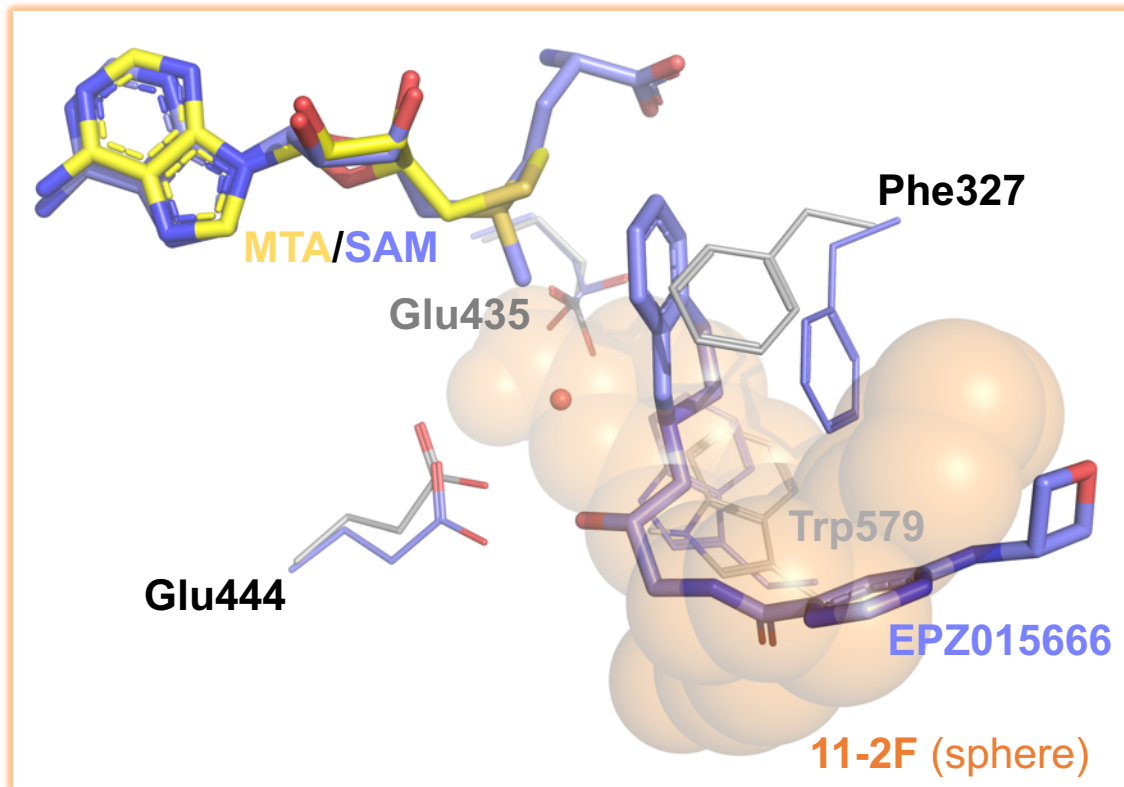


Structure comparison of apo- and holo- forms of PRMT5:MEP50



Structure alignment of bound-forms of PRMT5:MEP50

[11-2F + MTA]-bound form (grey) is aligned with [EPZ015666 + SAM]-bound form (purple)



EPZ015666 (IC₅₀ = 21.7 ± 5.0 nM)

11-2F ($IC_{50} = 0.7 \pm 0.2 \mu M$)

In [11-2F + MTA]-bound form, 11-2F:

- kicks out the **H₂O** molecule in EPZ015666-bound form by forming H-bonds with **Glu435** and **Glu444**,
- forms π - π interactions with **Phe327** and **Trp579**.

11-2F as a substrate-competitive inhibitor:

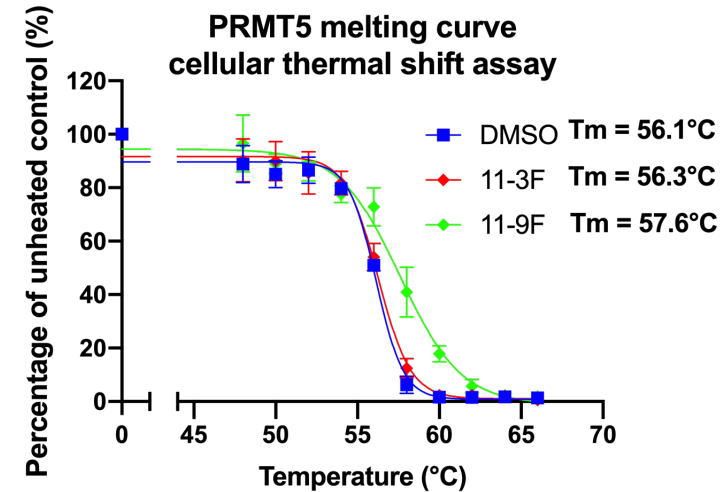
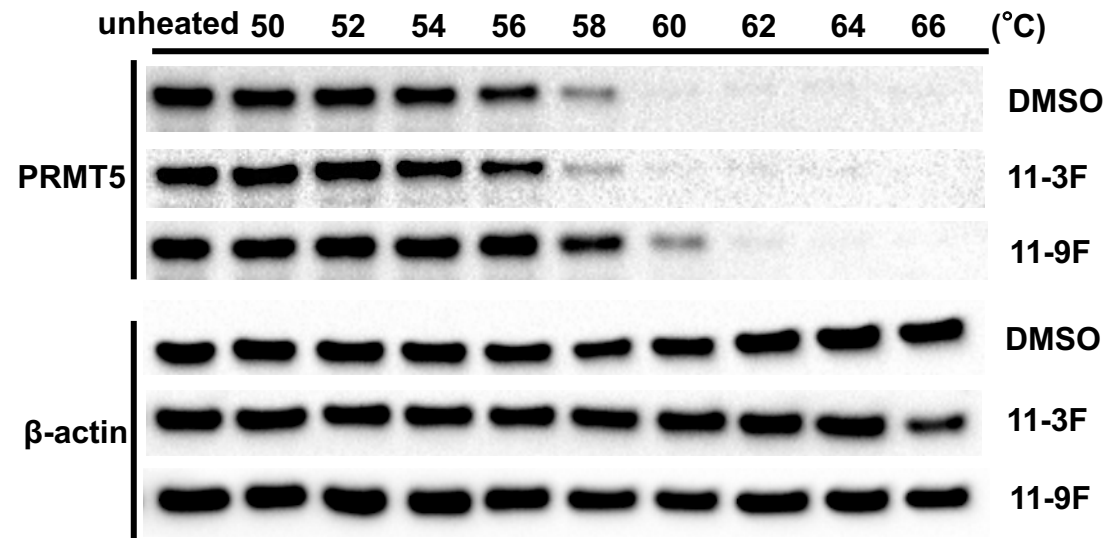
- does not rely on the pre-binding of SAM
- desired MOI in MTA-accumulated MTAP-deleted cancers

Target engagement study by CETSA

Cellular Thermal Shift Assay (CETSA)

11-3F ($IC_{50} > 500 \mu M$)

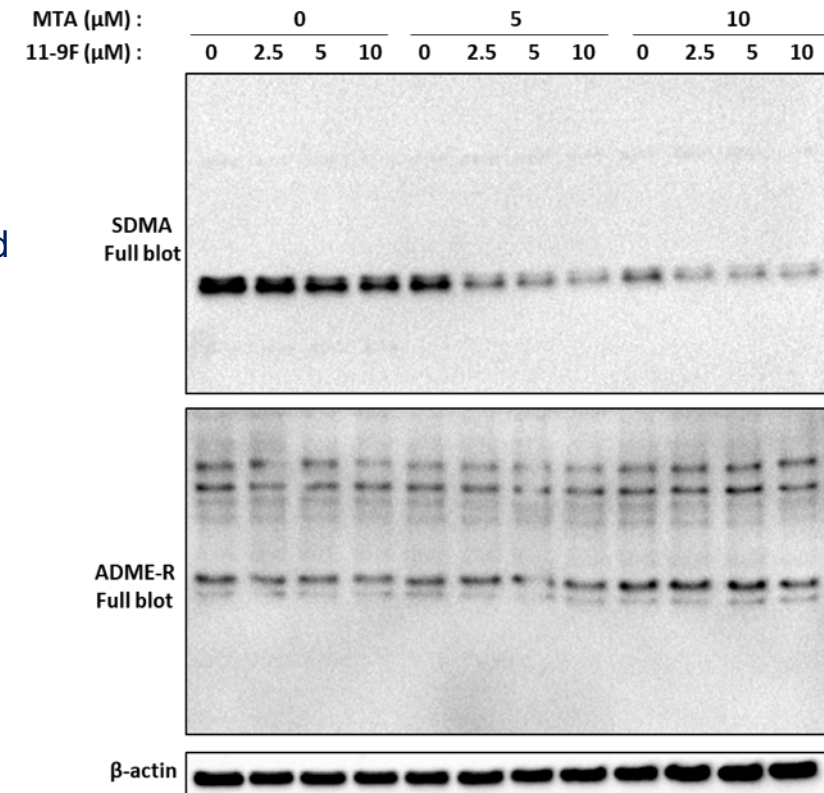
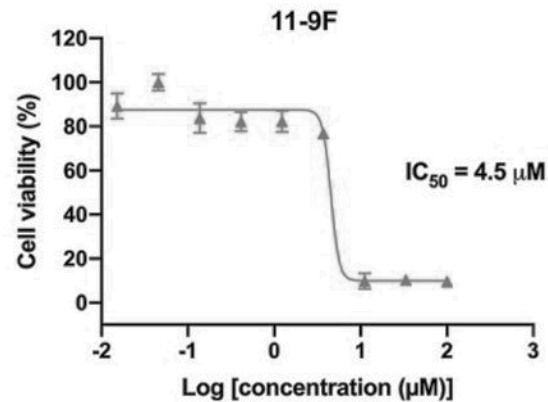
11-9F ($IC_{50} = 179.5 \pm 10.4 \text{ nM}$)



Protein methylation inhibition and selectivity in cells by western blot

Cellular effects of 11-9F in MDA-MB-231 breast cancer cell line (96 h)

- ❖ Concentration-dependent inhibition of cellular protein arginine symmetric dimethylation (**SDMA**) level was shown by western blot.
- ❖ MTA improved the inhibition activity of 11-9F, suggesting a synergistic effect.
- ❖ Unaffected protein arginine asymmetric dimethylation level (**ADME-R**) indicated a decent selectivity of 11-9F against Type I PRMTs.
- ❖ 11-9F showed anti-proliferation effect in MTT assay.



CONCLUSIONS



- ✓ The potency of designed inhibitors has been improved by more than 3000-fold.
- ✓ Designed inhibitors show a substrate-competitive mode of action in enzymatic assays.
- ✓ 11-9F binds to both apo- and cofactor-bound forms of PRMT5:MEP50 and shows binding preference to MTA-bound form.
- ✓ The flexible loop region at the active site of PRMT5 undergoes a huge conformational change upon cofactor binding.
- ✓ Interaction between 11-2F and PRMT5 is different from current inhibitors, making it a novel series of PRMT5 inhibitor.
- ✓ Treatment of 11-9F in combination with MTA improves its inhibition activity in cellular assays.

CONCLUSIONS



11-9F is a potent and selective PRMT5 inhibitor that binds to PRMT5 at substrate-binding site and shows synergistic effect with MTA *in vitro*.

FUTURE DIRECTION

- Compound optimization (potency↑, toxicity↓)
- Inhibition selectivity against other protein methyltransferases
- *In vitro* cellular assays to further investigate the anti-cancer effect of designed inhibitors
- Compound activity profile in cancer cell lines with different MTAP-expression levels
- *In vivo* evaluations in mice models

ACKNOWLEDGEMENTS

Li Lab Members:

Xiaozhi Yang (medicinal chemistry)
 Mohammad Rezaei, PhD
 Daniel Schultz
 Chen Zhou
 Gustavo Seabra, PhD
 Iram Hyder, PhD
 Dejun Li, PhD
 Chenglong Li, PhD (PI)

Previous Members:

Guqin Shi, PhD (computation)
 Linsen Li, PhD (medicinal chemistry)
 Hongshan Lai (medicinal chemistry)

Jiang Lab Members:

Gaya Yadav, PhD (cryo-EM)
 Qiu-Xing Jiang, PhD (PI)

Committee Members:

Prof. Chenglong Li (Chair)
 Prof. Qiu-Xing Jiang
 Prof. Robert McKenna
 Prof. Jianrong Lu
 Prof. Hendrik Luesch

UF Center for Structural Biology:

Mavis Agbandje-McKenna, PhD

Cryo-EM Data Collection:

UF Interdisciplinary Center for
 Biotechnology Research
 UNIVERSITY of FLORIDA



NIH NATIONAL CANCER INSTITUTE
 NCI at Frederick
 National Cryo-Electron Microscopy Facility

Funding:

NIH/NCI R01CA212403
 UFHCC Shands fund
 NIH (R21GM131231,
 R01GM111367, R01GM093271,
 1U24GM116788)
 CF Foundation (JIANG15G)
 ROF2019 pilot award from UF



UF Department of Biochemistry
 and Molecular Biology
 College of Medicine
 UNIVERSITY of FLORIDA

UF College of Pharmacy
 UNIVERSITY of FLORIDA

UFHealth
 CANCER CENTER